



Spatial integration of antimicrobial peptides controls release and activity in oral delivery systems

Cesar Augusto Roque-Borda, Ph.D.



CRS2026
ANNUAL MEETING & EXPOSITION
LISBON
— PORTUGAL —

MULTISECTOR
INNOVATION
for ADVANCING
DELIVERY SCIENCE

JULY 6–9, 2026

Why oral AMPs fail

- ☐ Proteolysis
- ☐ Acid degradation
- ☐ Burst release
- ☐ Poor intestinal targeting
- ☐ Electrostatic destabilization
- ☐ Loss of bioactivity before reaching site

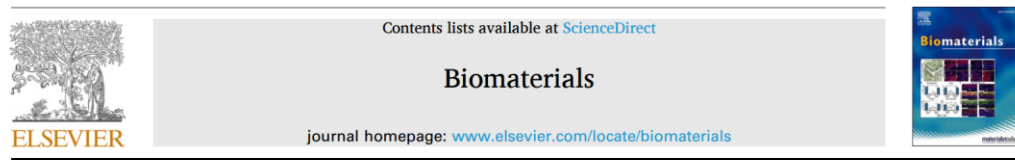
**Most delivery systems
treat AMPs as passive
cargo.**

Aim:

We investigated whether the AMP itself could become a structural component of the matrix.



Strategy A — Protected encapsulation (core-shell double coating)



Chitosan and HPMCAS double-coating as protective systems for alginate microparticles loaded with Ctx(Ile²¹)-Ha antimicrobial peptide to prevent intestinal infections

Cesar Augusto Roque-Borda^{a,*}, Mauro de Mesquita Souza Saraiva^b,
Wagner Dias Macedo Junior^c, José Carlos Estanislao Márquez Montesinos^d,
Andréia Bagliotti Meneguín^a, Anna Beatriz Toledo Borges^{a,1}, Edson Crusca Junior^e,
Saulo Santesso Garrido^e, Adriana Maria de Almeida^b, Reinaldo Marchetto^e, Marlus Chorilli^a,
Angelo Berchieri Junior^b, Silvio Rainho Teixeira^c, Fernando Rogério Pavan^a,
Eduardo Festozo Vicente^{f,*}



Strategy B — Electrostatic peptide integration (self-assembled nano-in-micro structures)

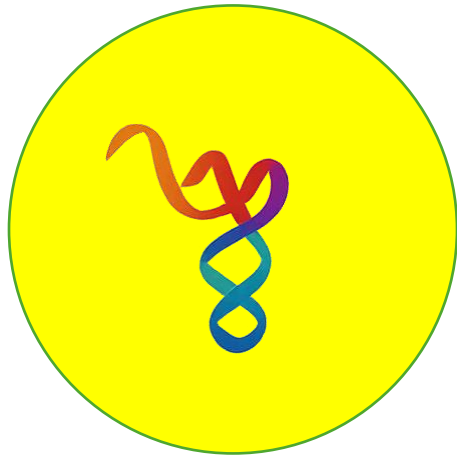


Alginate-pectin microparticles embedding self-assembling antimicrobial peptides and resveratrol for antimicrobial and anti-inflammatory applications

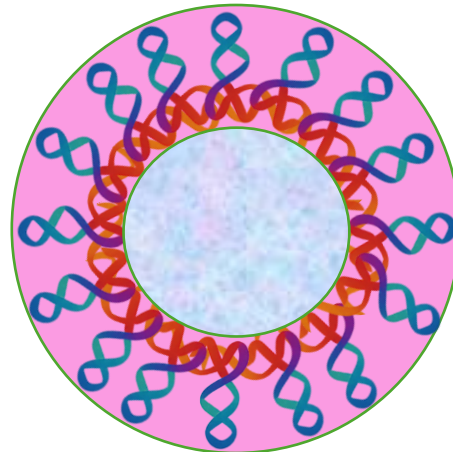
Cesar Augusto Roque-Borda^{a,*}, Marco Roberto Chávez-Morán^b,
Laura Maria Duran Gleriani Primo^c, José C.E. Márquez Montesinos^{d,i},
Vinicius Martinho Borges Cardoso^c, Mauro M.S. Saraiva^e, Caroline Maria Marcos^c,
Marlus Chorilli^c, Fernando Albericio^{f,g,h,*}, Beatriz G. de la Torre^h, Fernando Rogério Pavan^c,
Andréia Bagliotti Meneguín^{c,***}



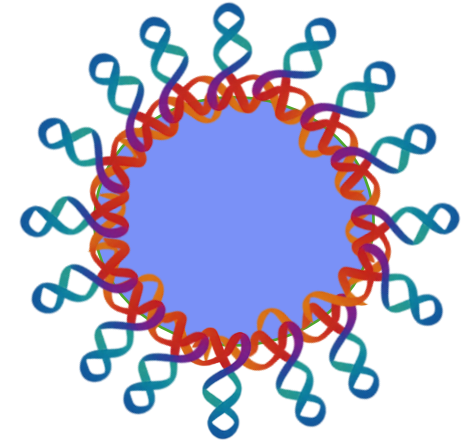
Micro/Nanoparticles



Core: Peptide



Core: Drug/Bioactive compound



Core: Drug/Bioactive compound

Knowing the systems

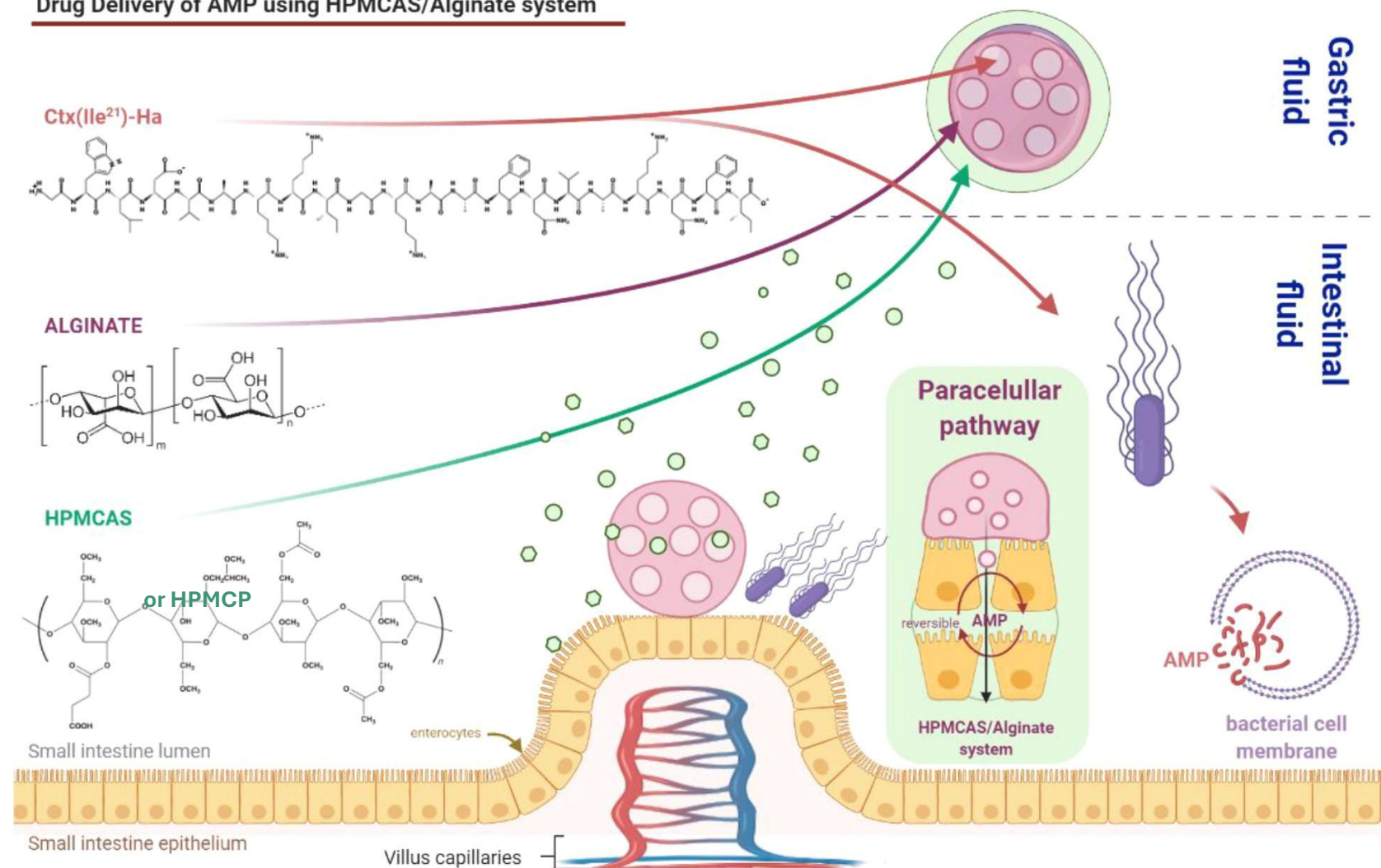


Alginate-based microparticles coated with HPMCP/AS cellulose-derivatives enable the Ctx(Ile²¹)-Ha antimicrobial peptide application as a feed additive

Cesar Augusto Roque-Borda^a, Hanyeny Raiely Leite Silva^a, Edson Crusca Junior^b, Jéssica Aparecida Serafim^c, Andréia Bagliotti Meneguim^d, Marlis Chorilli^d, Wagner Costa Macedo^e, Silvio Rainho Teixeira^e, Elisabete Aparecida Lopes Guastalli^f, Nilce Maria Soares^f, Jessica M.A. Blair^g, Zoe Pikramenou^h, Eduardo Festozo Vicente^{c,*}

- Alginate = ionic gelation
- Chitosan = mucoadhesion
- HPMCAS/HPMCP = enteric protection

Drug Delivery of AMP using HPMCAS/Alginate system

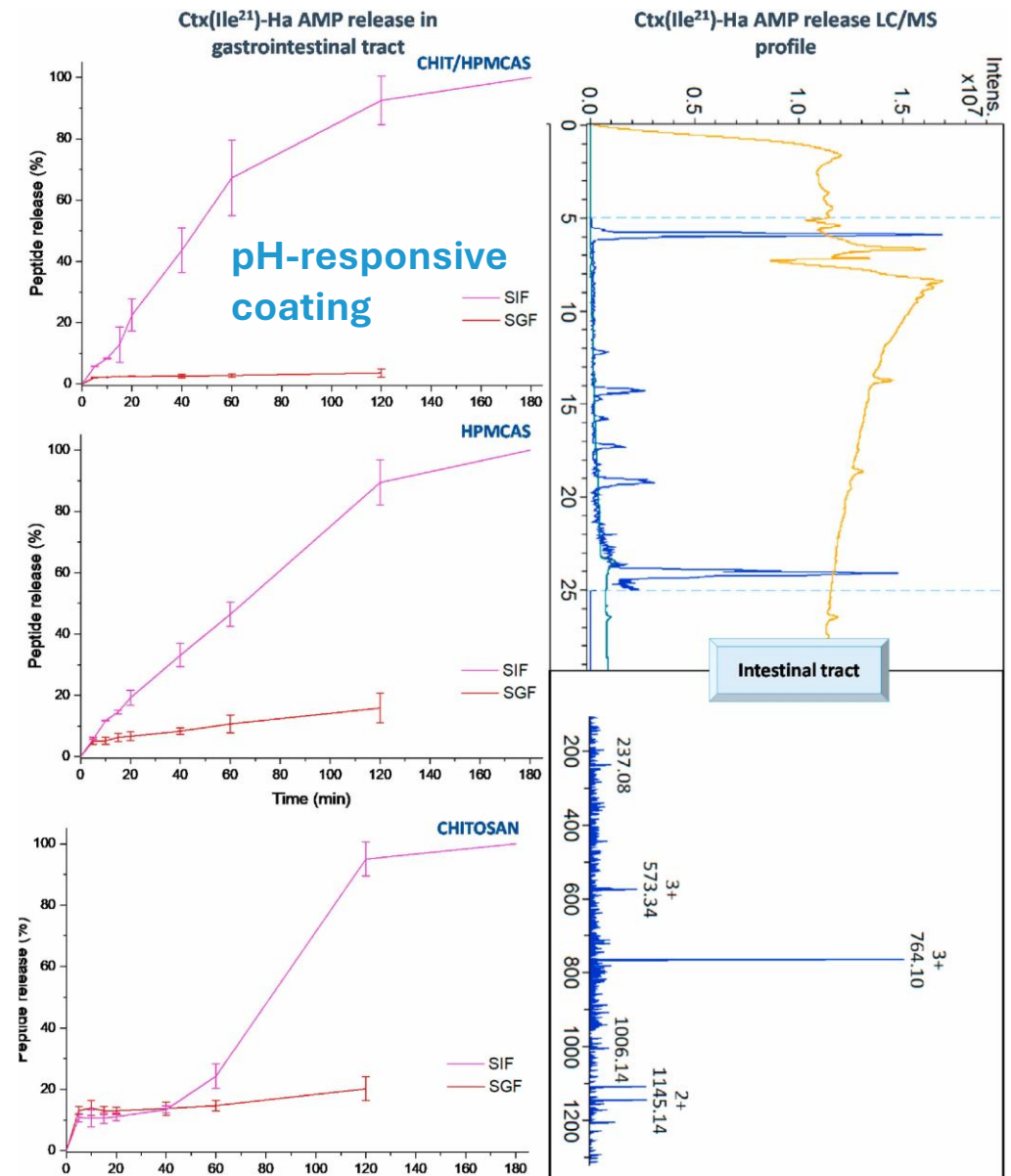


Why double coating?

- protect AMP in gastric environment
- release in intestine
- low release at gastric pH
- higher release at intestinal pH

intestinal-
triggered release

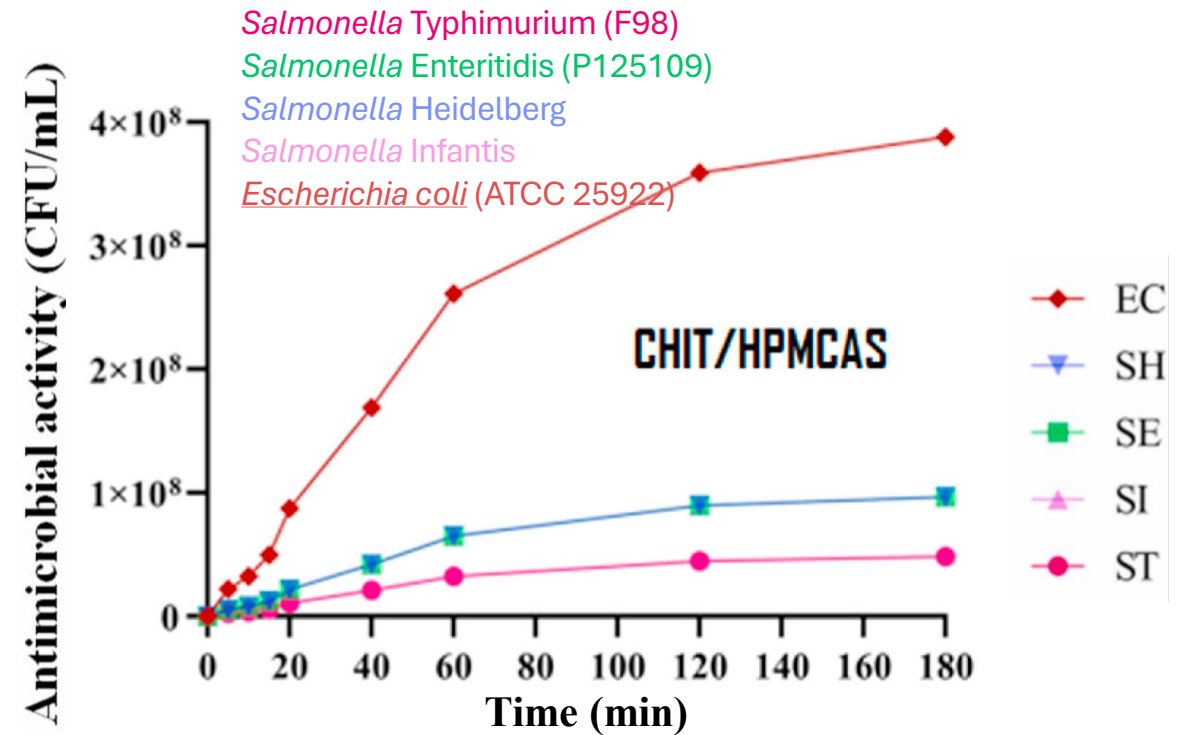
The system delayed
premature peptide
exposure.



Biological outcome

- ❑ antimicrobial activity preserved
- ❑ peptide survives GI simulation
- ❑ intestinal pathogen control

Encapsulation preserved
functionality but limited immediate
interaction with the environment.

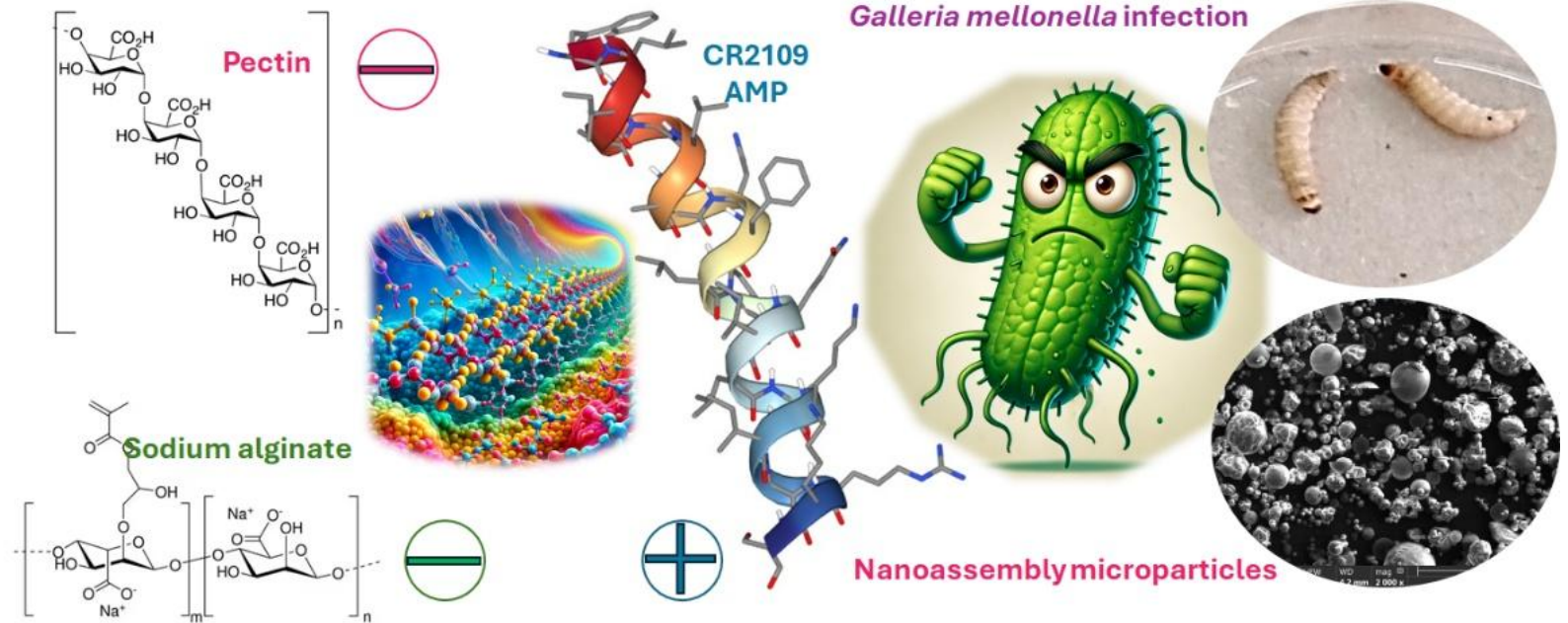


What if the peptide is not only protected... but structurally integrated?

Electrostatic self-assembly concept

- ☐ cationic peptide
- ☐ anionic alginate/pectin
- ☐ spontaneous nanoassembly

Peptide became part of matrix organization

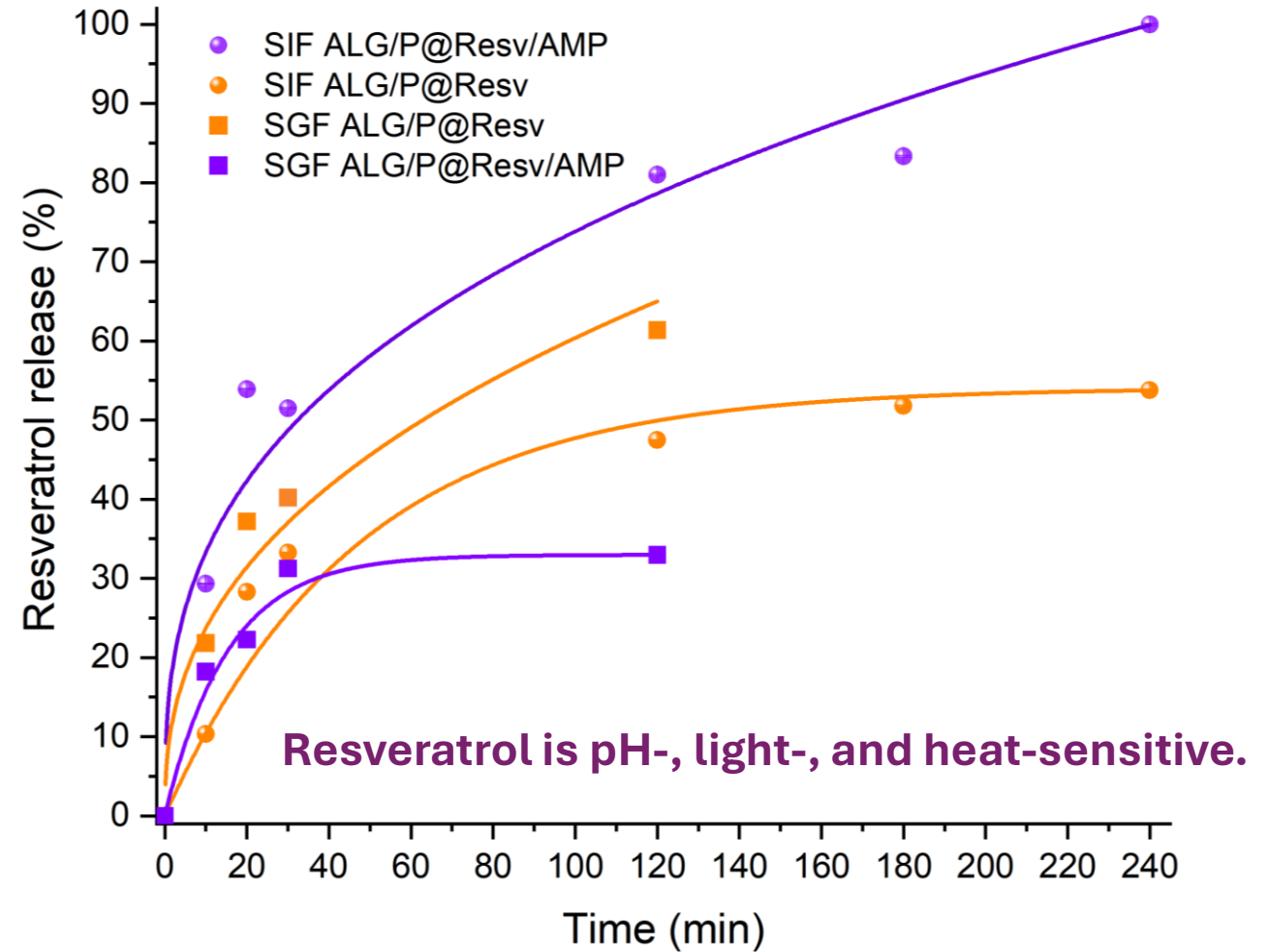


Self-assembly mechanism

Possible driving forces

- ☐ electrostatic interactions
- ☐ hydrophobic interactions
- ☐ hydrogen bonding
- ☐ polymer chain rearrangement

The AMP altered diffusional behavior of the matrix.



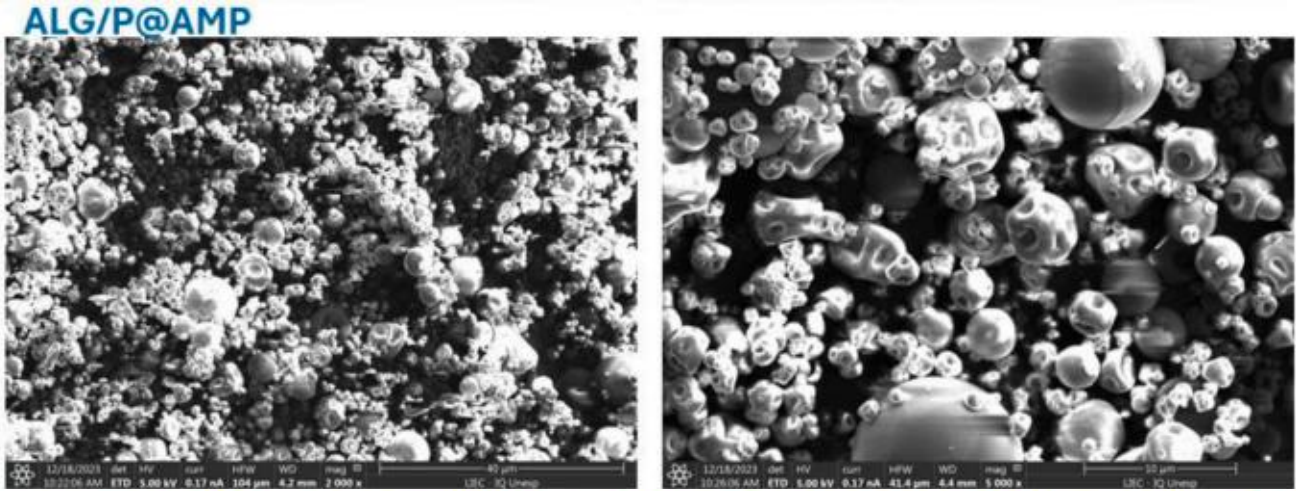
Microparticle architecture

The peptide reorganized the colloidal environment before spray drying

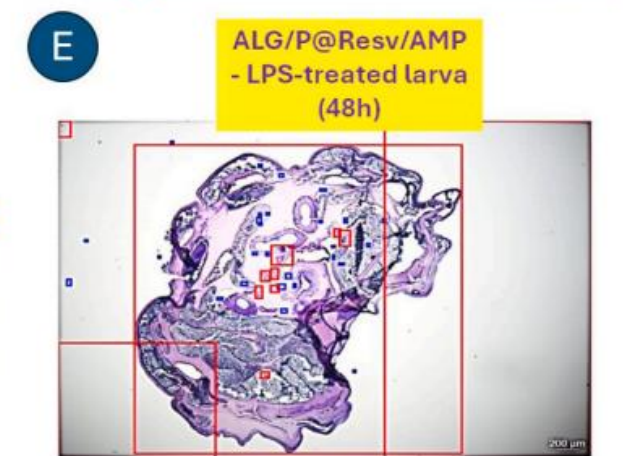
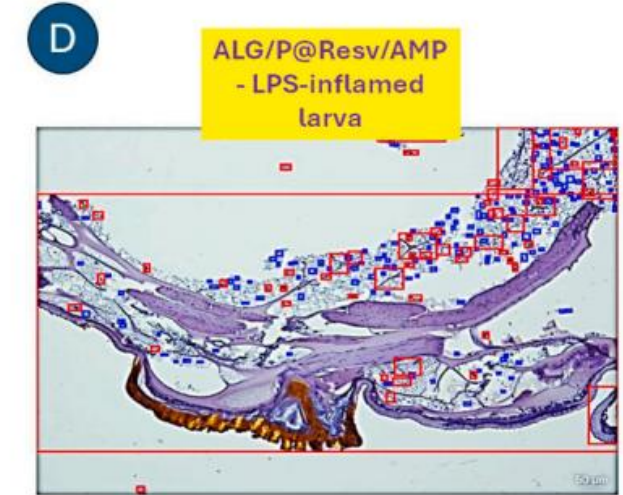
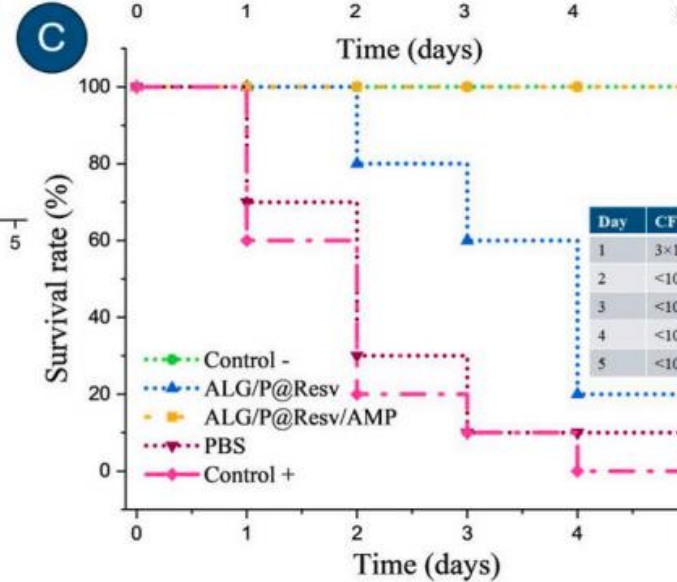
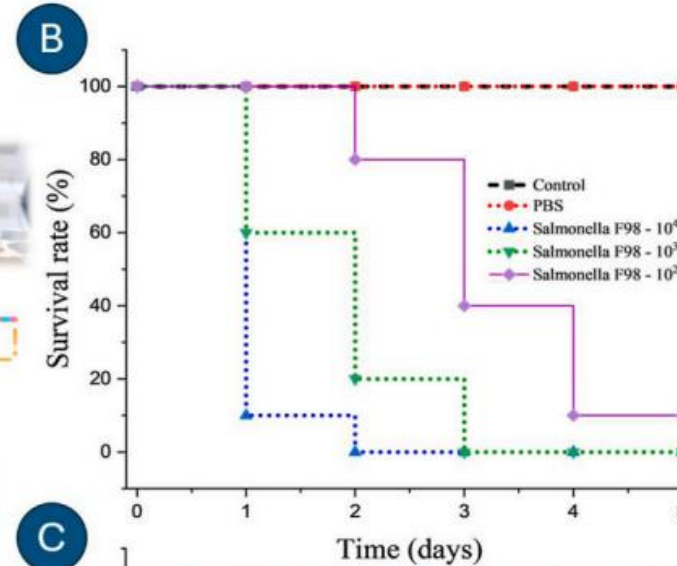
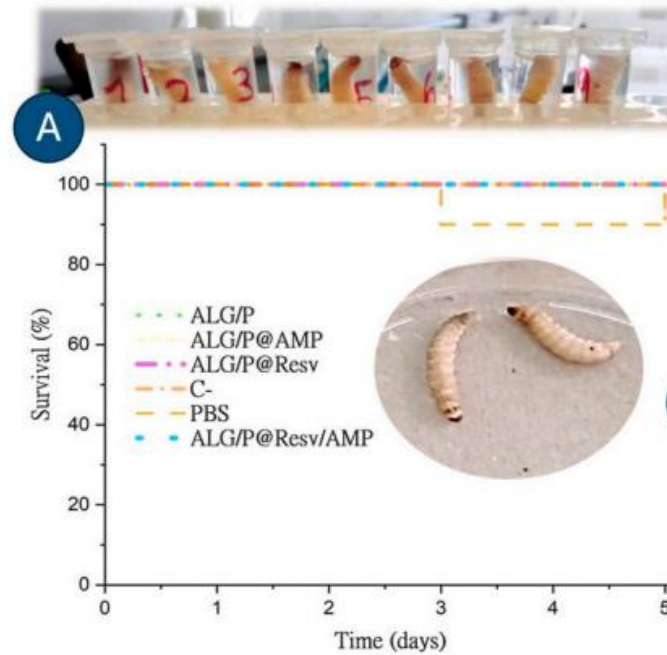
Table 2
Characteristics of nano-in-microparticles formed with peptide and resveratrol.

System/dispersion	EE (%)		DC (%)		Yield (%)	PDI	Zeta potential (mV)	Size (nm)
	Resv	AMP	Resv	AMP				
Blank	–	–	–	–	33.22	0.552	–25.0	894.5
ALG/P@Resv	78.21	–	39.10	–	40.87	0.456	–17.5	362.7
ALG/P@AMP	–	66.39	–	0.259	14.44	0.364	–7.4	220.6
ALG/P@Resv/AMP	76.11	72.84	89.23	2.73	4.06	0.185	–18.2	152.8
AMP	–	–	–	–	–	0.338	+10.6	–
Resv	–	–	–	–	–	0.570	–5.8	880.2

Zeta potential shifts suggested peptide–polymer nanoassembly prior to spray drying



Anti-inflammatory and antimicrobial effects



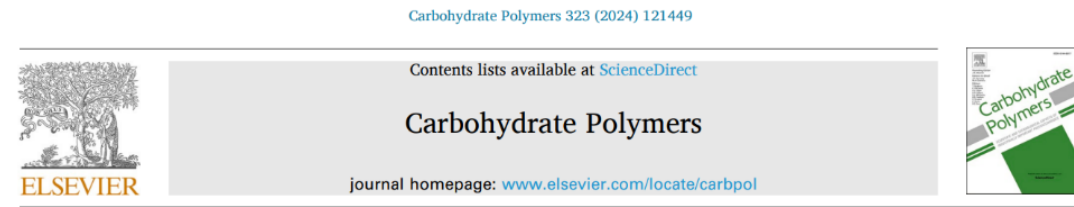
- ☐ sustained activity
- ☐ prolonged modulation
- ☐ in vivo reduction of inflammation
- ☐ bacterial load reduction



A third possibility: surface-exposed AMPs

Why graft the peptide?

The AMP no longer behaves as cargo, but as a functional nanointerface.



Antimicrobial peptides grafted onto the surface of *N*-acetylcysteine-chitosan nanoparticles can revitalize drugs against clinical isolates of *Mycobacterium tuberculosis*

Laura Maria Duran Gleriani Primo^{a,1}, Cesar Augusto Roque-Borda^{a,h,*},
Christian Shleider Carnero Canales^b, Icaro Putinhon Caruso^c, Isabella Ottenio de Lourenço^c,
Vitória Maria Medalha Colturato^d, Rafael Miguel Sábio^e, Fernando Alves de Melo^c,
Eduardo Festozo Vicente^f, Marlus Chorilli^e, Hernane da Silva Barud^d, Paula Aboud Barbugli^g,
Henrik Franzyk^h, Paul Robert Hansen^h, Fernando Rogério Pavan^{a,*}



Multiple attack concept

NP core:

- ❑rifampicin delivery

Chitosan:

- ❑cationic destabilization

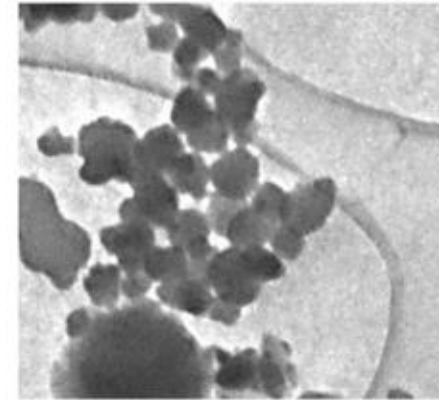
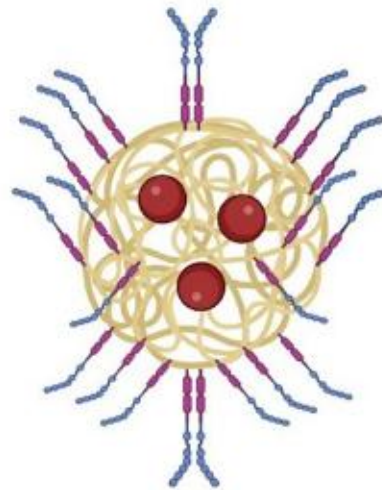
NAC:

- ❑biofilm-related modulation

Surface AMP:

- ❑membrane interaction
- ❑receptor interaction
- ❑possible efflux modulation

Bionanoconjugate system are active
against resistant *Mycobacterium*
tuberculosis



MDR/XDR *Mycobacterium*
tuberculosis



N-acetylcysteine



Ctx(Ile21)-Ha-Ahx-Cys



Chitosan



Rifampicin



Mycobacterium tuberculosis

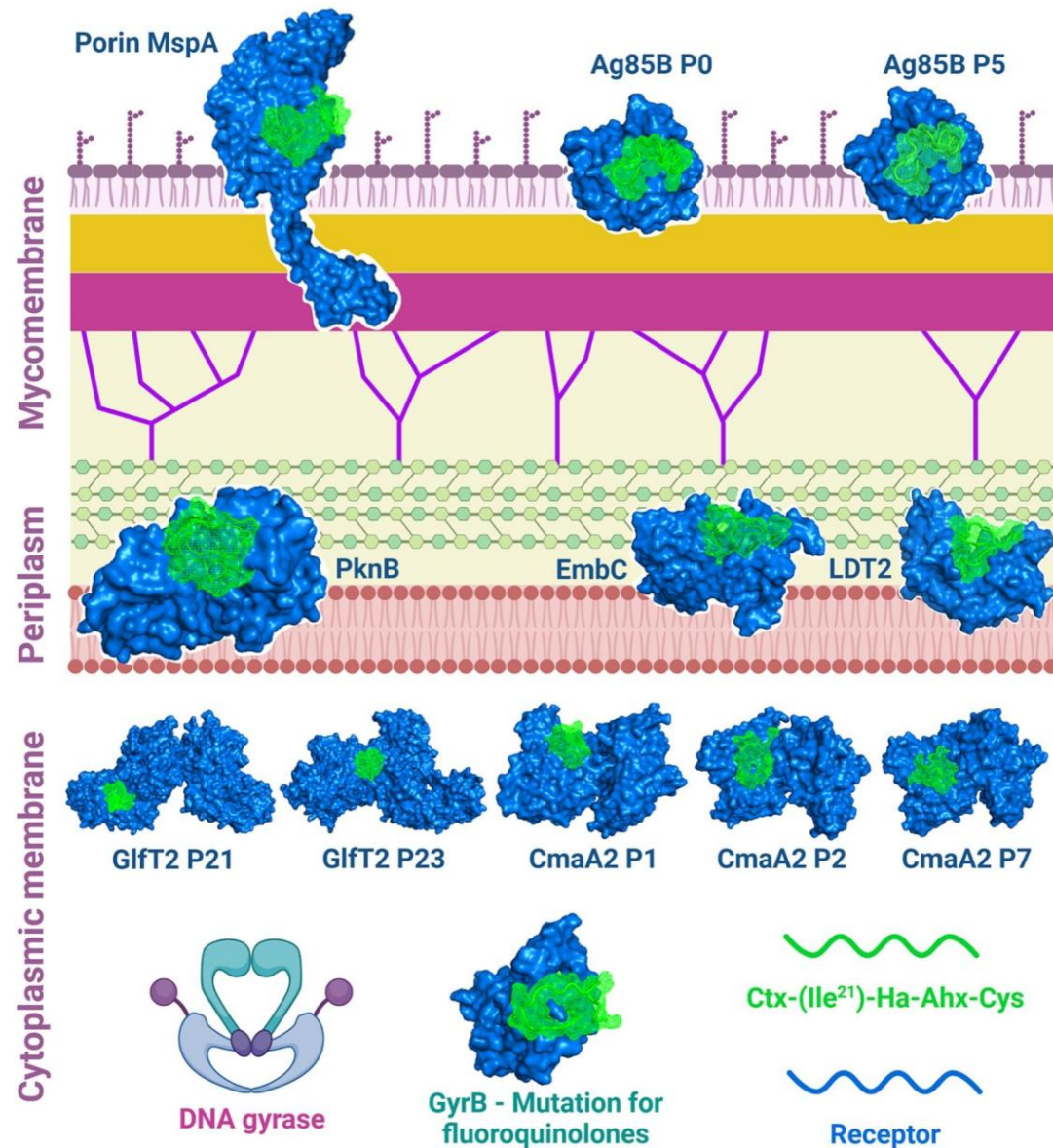


Table 3

MIC values of antibiotics and NP formulations by the REMA method after 7 days of exposure.

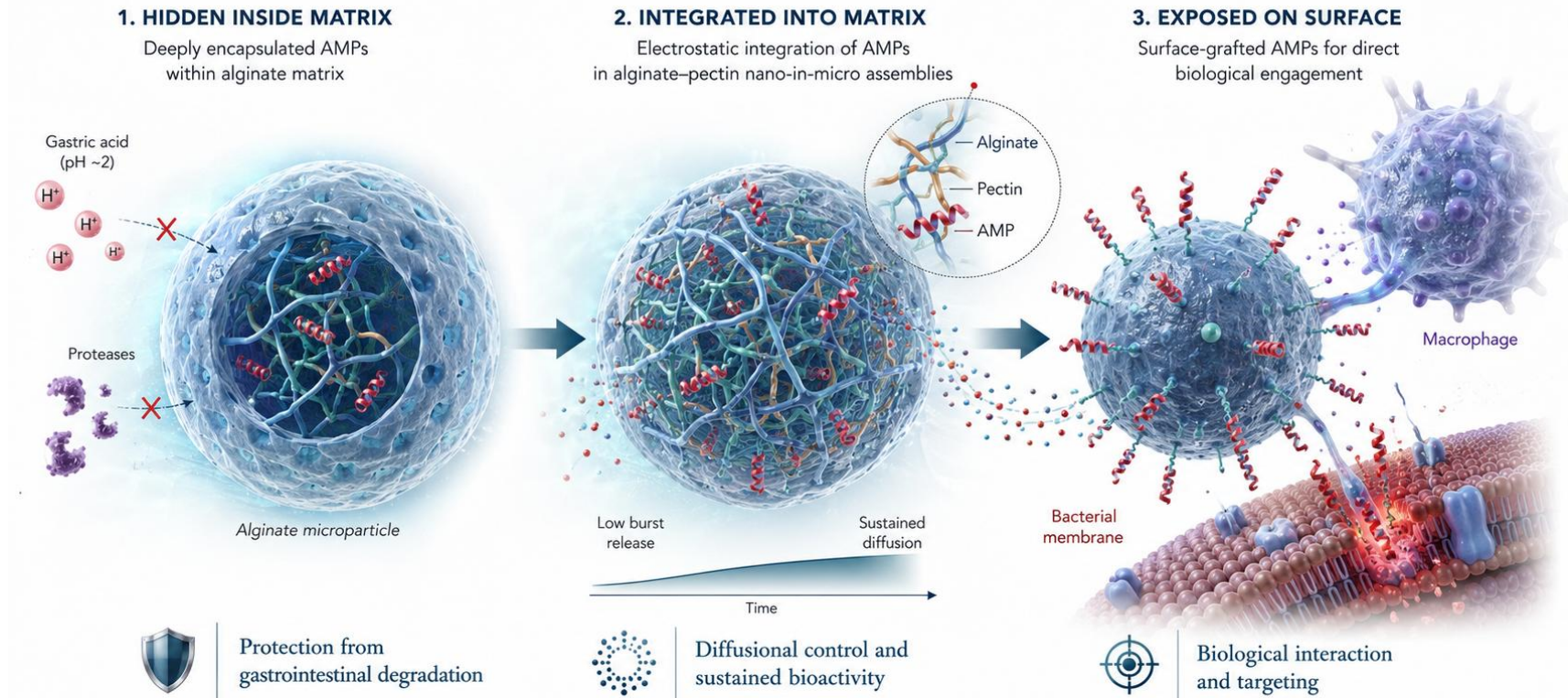
Code	Formulation	H37RV (µg/mL)	CF 169 (µg/mL)	CF 110 (µg/mL)
INH	Isoniazid	0.5–1.0	>50	>50
RIF	Rifampicin	0.5	>25	16.29
GAT	Gatifloxacin	0.03–0.25	1.2	<0.05
MOX	Moxifloxacin	0.03–0.5	1.6	<0.05
OFL	Ofloxacin	0.71	11.5	0.3
AMK	Amikacin	0.5–1.0	0.6	0.5
CAP	Capreomycin	2.0	1.4	3.0
KAN	Kanamycin	2.0	1.5	2.8
NP1	NAC-CS	NA	NA	NA
NP2	CS@RIF ^a	<0.977	NA	NA
NP3	Ctx-NAC-CS	NA	NA	NA
NP4	NAC-CS@RIF ^a	1.47	199.05	NA
NP5	CS	NA	NA	NA
NP6	Ctx-CS	NA	NA	NA
NP7	Ctx-CS@RIF ^a	<0.977	104.34	NA
NP8	Ctx-NAC-CS@RIF ^a	<0.977	0.977	<0.977
Ctx	Ctx(Ile ²¹)-Ha-Ahx-Cys	7.57	12.24	>250
Ctx wild-type	Ctx(Ile ²¹)-Ha	14.0	NT	NT
NAC	N-acetylcysteine	NA	NA	NA

NA: no activity. NT: not tested.

CF169 strain: XDR MTB. CF110 strain: MDR MTB.

^a MIC values based on RIF concentration. All NPs were tested based on their weight.

Conceptual convergence



❑ Peptide localization matters

❑ Matrix interactions shape bioactivity— *Spatial positioning of AMPs may be as important as peptide sequence itself.*

❑ Electrostatic integration alters diffusion

❑ Self-assembly modulates biological output

❑ Nano-in-micro systems enable sustained AMP action

❑ AMP-matrix interactions are an overlooked design parameter



CRITICAL ANALYSIS IN PEPTIDE CHEMISTRY

Year in review

nature reviews chemistry

Peptide chemistry

<https://doi.org/10.1038/s41570-025-00786-4>

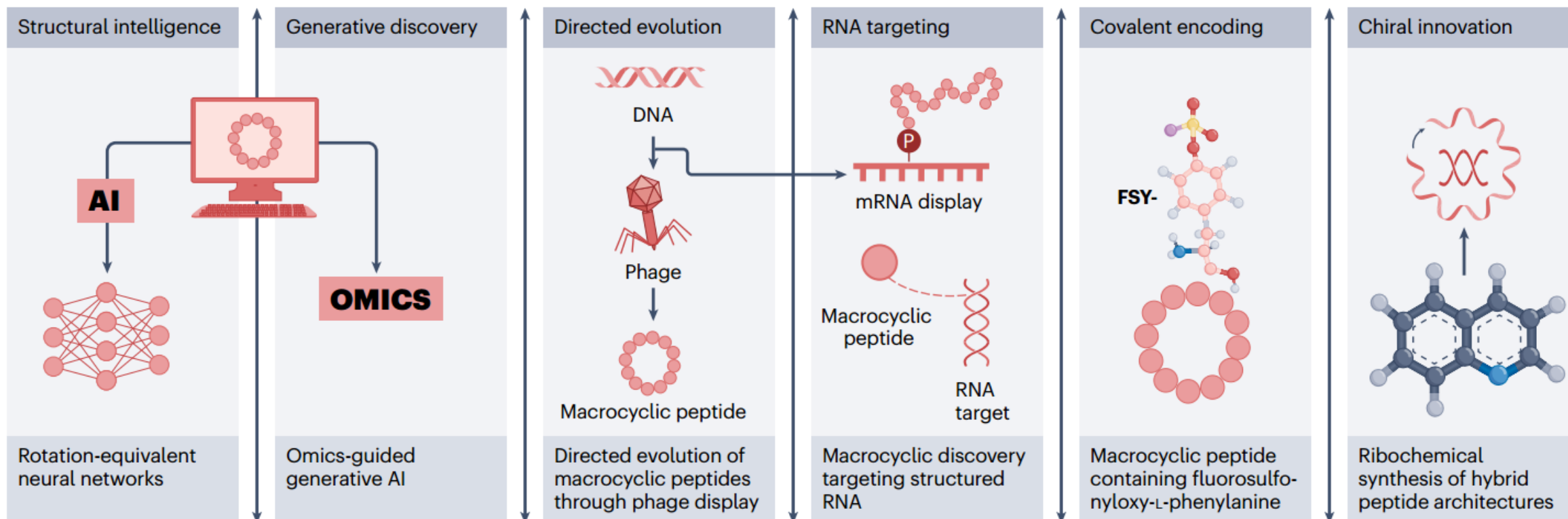
Ref:

Redefining peptide chemistry beyond
accumulating analogues

Cesar Augusto Roque-Borda, Fernando Rogério Pavan, Beatriz G. de la Torre
& Fernando Albericio

Check for updates

Peptide design enters the generative age



Acknowledgments

Thank You For Your
Attention

ORCID



E-mail: cesar.roque@unesp.br or cesarb@edu.ulisboa.pt