

Tech Session 4: Nanomedicine and Nanoscale Delivery

Bruno Silva



CRS 2023 ANNUAL MEETING & EXPOSITION

JULY 24-28, 2023 **Paris Hotel** » **Las Vegas, NV, USA**

THE FUTURE OF DELIVERY SCIENCE

Willkommen
Welcome
Bienvenue



A Criterion for Predicting Stable Assembly of Lipid-Polymer-Nucleic Acid Nanoparticles

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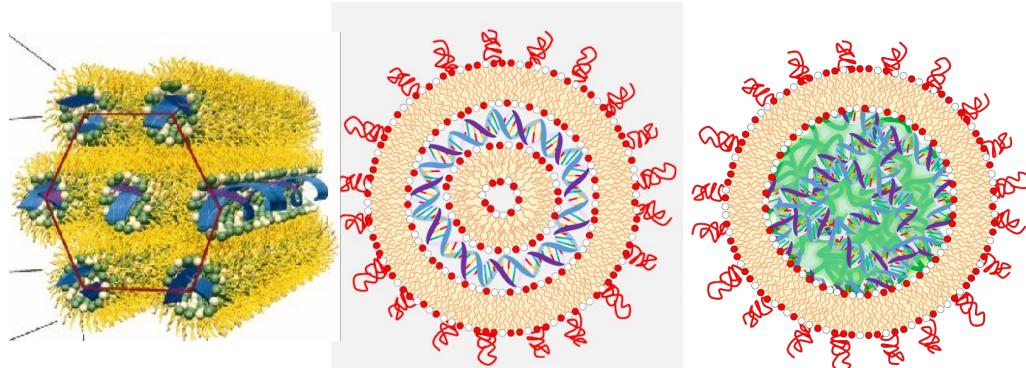


Bio-Nano Assembly group @ Empa

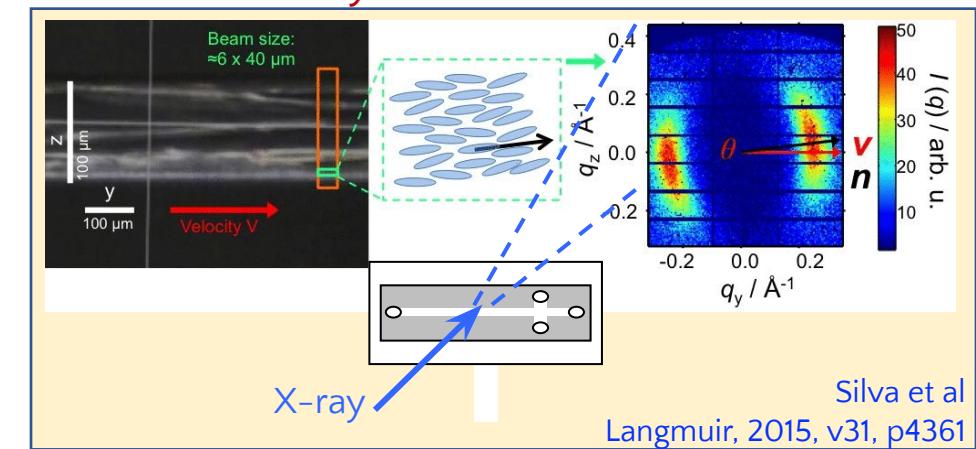
We develop new bio-nano-inspired nanosystems for health,
and **small-angle scattering (and beyond)** methods & concepts to understand them

Emphasis: on nonviral systems for gene therapeutics

- DNA-Lipid complexes, LCs & NPs

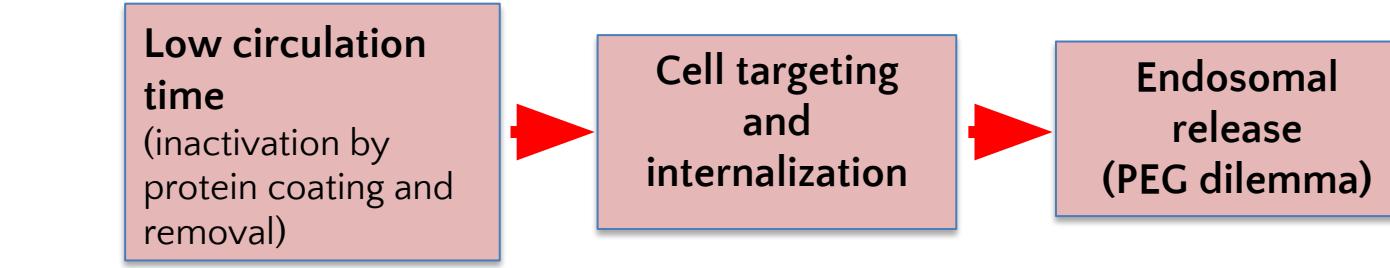


- Microfluidics for directed assembly
- in-situ X-ray



Complex designs to overcome complex tasks

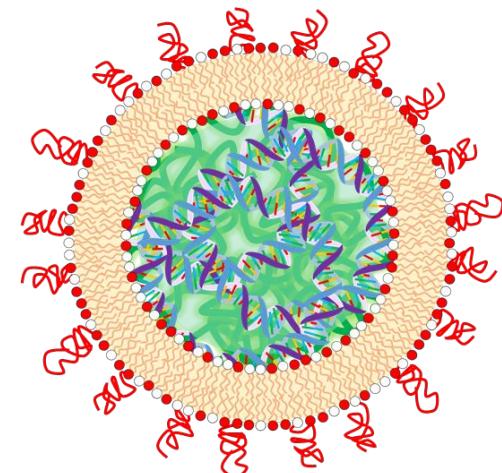
A successful gene-delivery nanosystems must overcome several biological barriers



Lipid Envelope

Targeting peptides
(RGD, KALA, etc)

Diffusible PEG coating

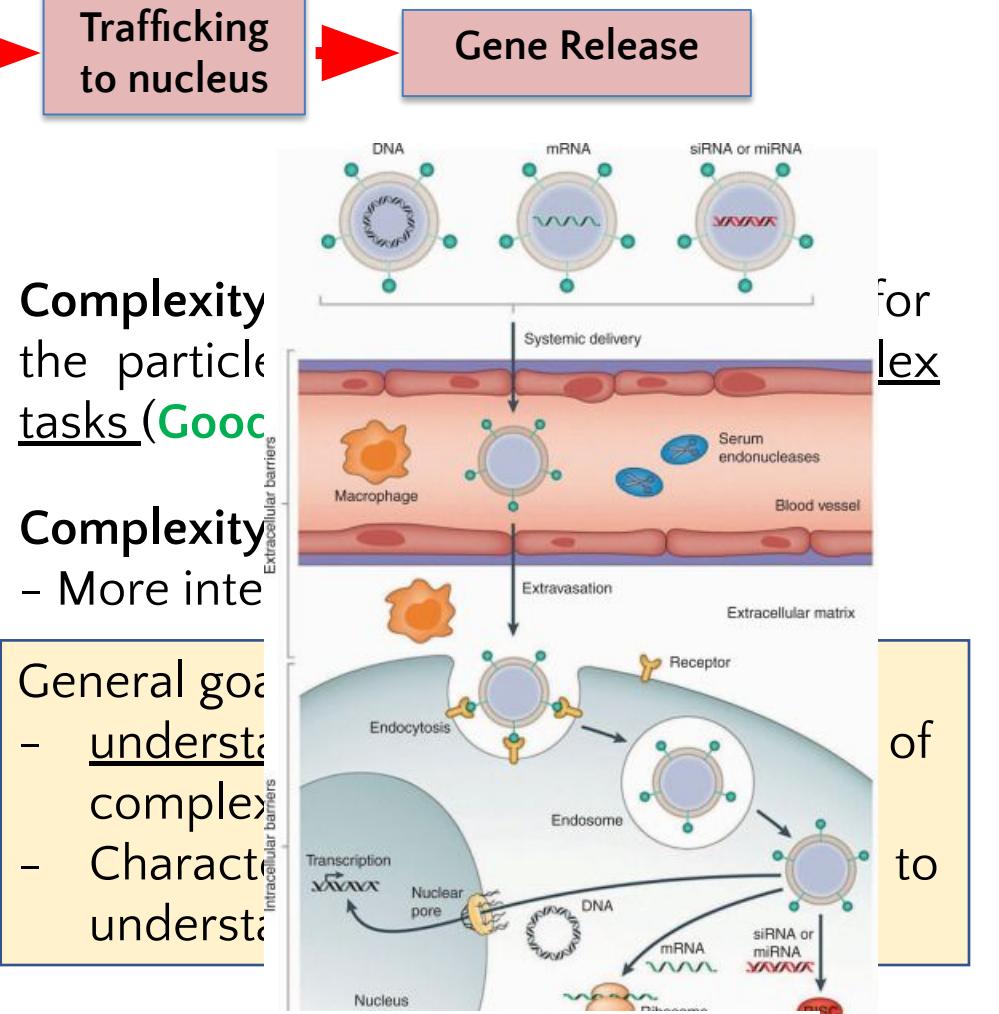


Lipid-polymer core shell NPs (LPNPs)

Polymer Core

NLS tags (SV40)

Destabilization for release
(e.g. disulfide bonds)

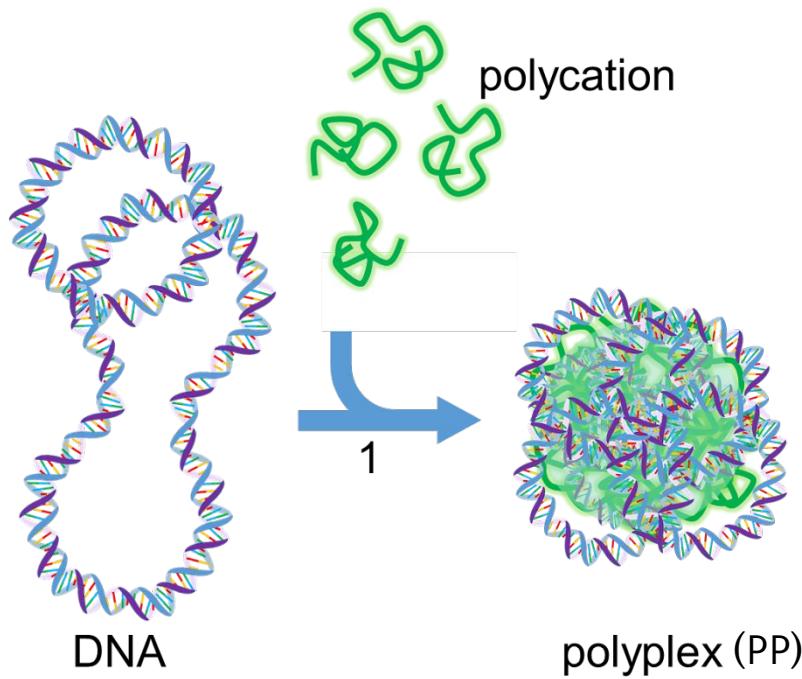


Complexity
the particle tasks ([Gooc](#))

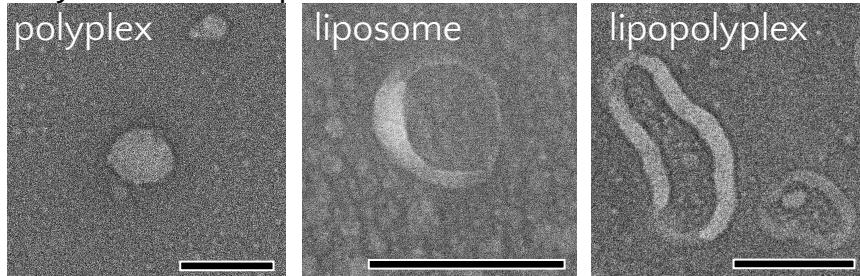
Complexity
- More int

General goal
- understan
complex
- Characte
understan

LPNPs: liposome-polycation-DNA nanoparticles



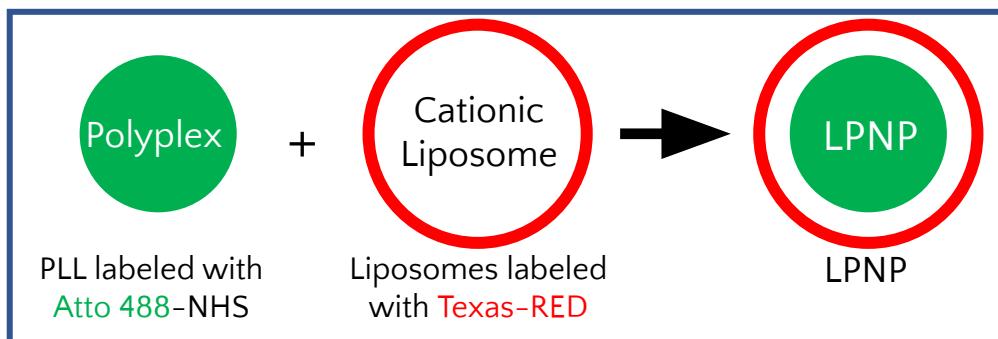
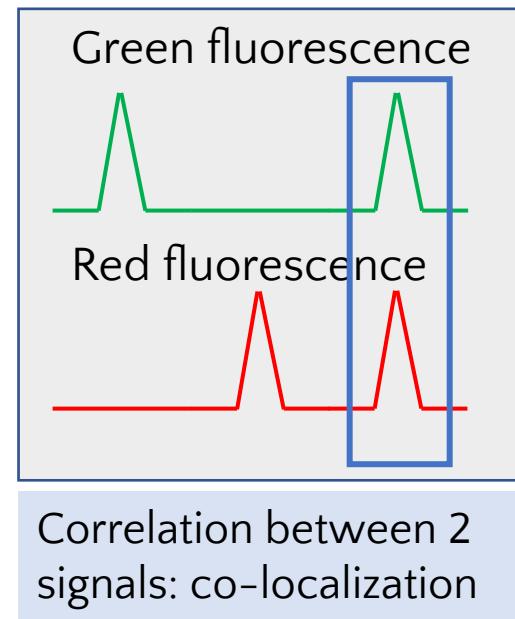
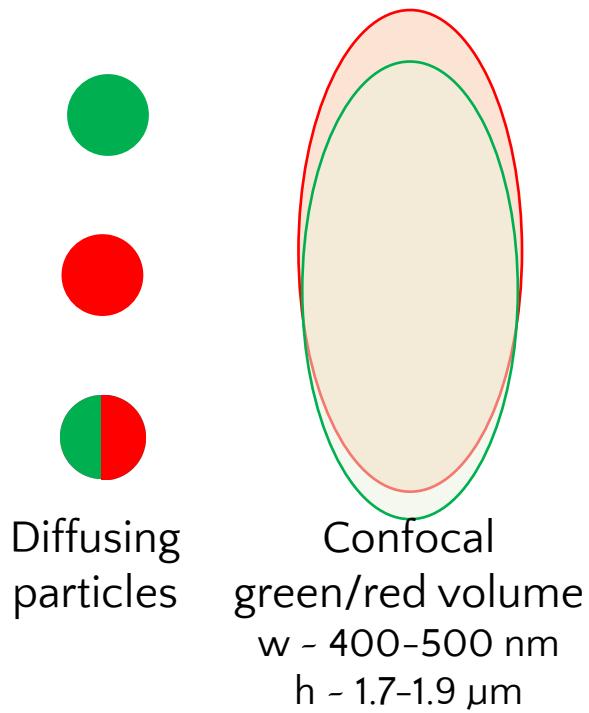
- 1) Charge inversion drives assembly – **Electrostatics** (identical to LbL)
- 2) For final particle positive charge, core should be negative (excess DNA)
 - Poly-L-lysine used as polycation (ubiquitous)
 - PEGylated Cat. Liposomes (80% DOTAP, 10% DOPC, 10% PEG)
- 3) Other pairwise interactions need to be considered
- 4) Liposomes may displace DNA from polycation
(c.f. Cat. Liposome – nucleosomes, [Berezhnay, Korolev, Nordenskiöld, Adv. Colloid Int. Sci. 2014](#))
- 5) Difficult to characterize with conventional methods



TEM methods reveal some details, but:

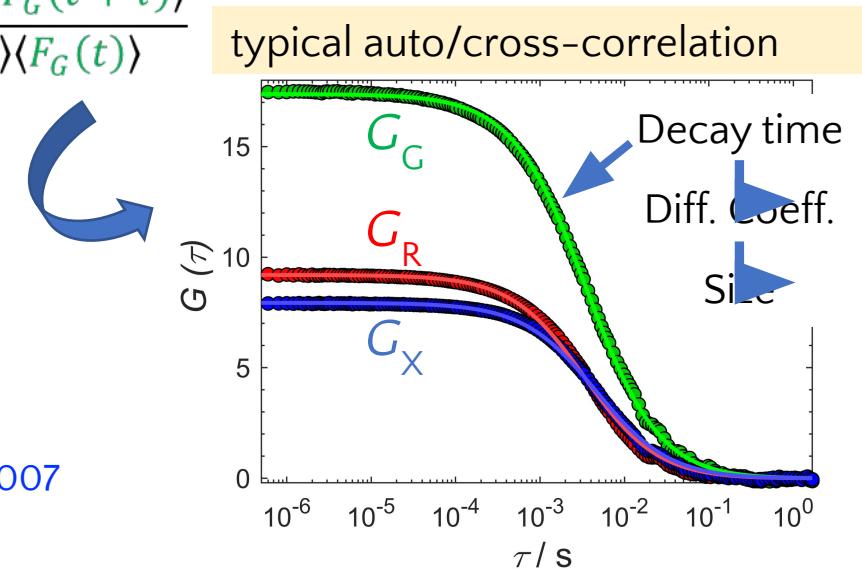
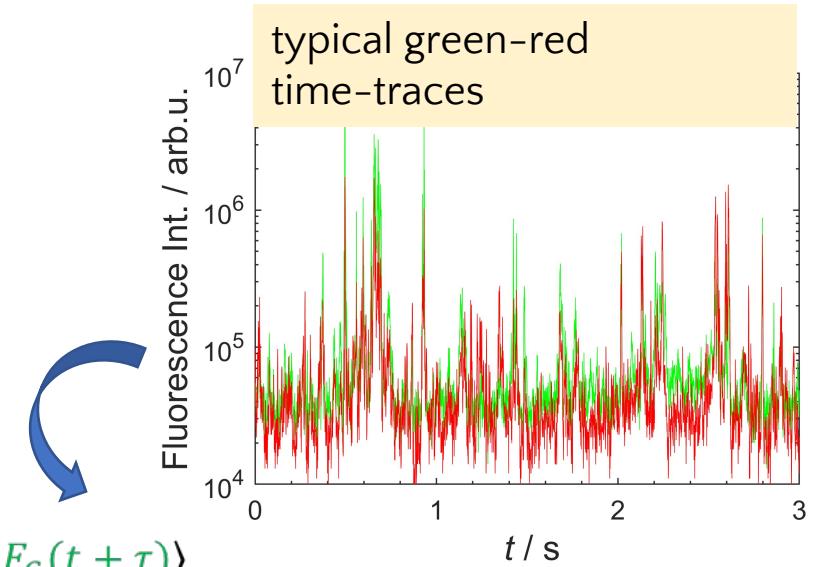
- How many particles have all three components?
- What is the optimal ratio between **Polymer**, DNA & **Lipid**?

FCCS - Fluorescence Cross-Correlation Spectroscopy



Bacia & Schwille, Nature Protocols, 2007

$$G_X(\tau) = \frac{\langle \delta F_R(t) \delta F_G(t + \tau) \rangle}{\langle F_R(t) \rangle \langle F_G(t) \rangle}$$



FCCS - Fluorescence Cross-Correlation Spectroscopy

Amplitudes of auto- and cross-correlation depend on particle concentration

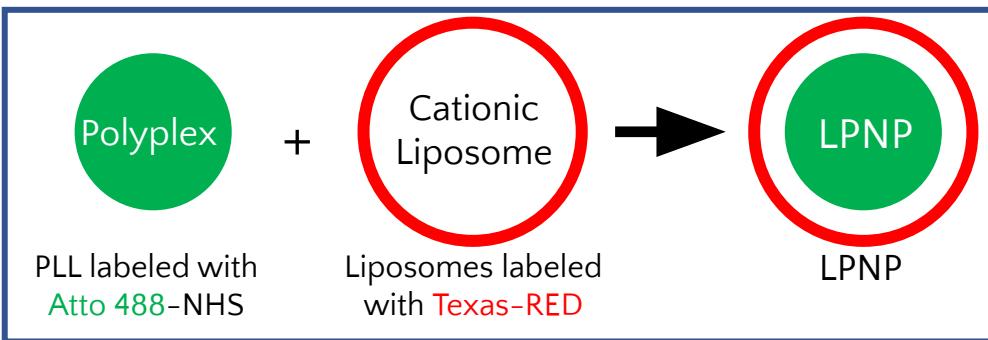
$$A_G = \sum_i \eta_{i,G}^2 N_{i,G} / \left(\sum_i \eta_{i,G} N_{i,G} \right)^2$$

$$A_R = \sum_i \eta_{i,R}^2 N_{i,R} / \left(\sum_i \eta_{i,R} N_{i,R} \right)^2$$

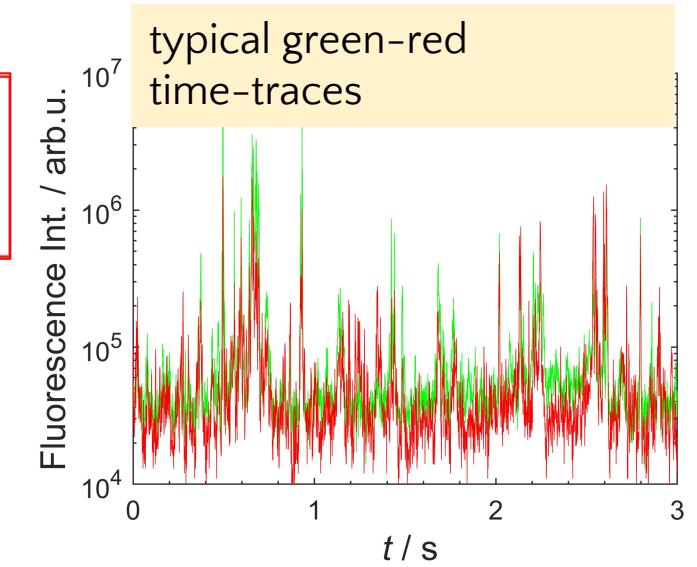
$$A_{RG} = \sum_i \eta_{i,G} \eta_{i,R} N_{i,RG} / \left[\left(\sum_i \eta_{i,G} N_{i,G} \right) \left(\sum_i \eta_{i,R} N_{i,R} \right) \right]$$

Auto-correlation A_G & $A_R \propto 1/C_i$ – inversely proportional to concentration

Cross-correlation $A_{RG} \propto C_{i,RG}$ – proportional to conc. of Co-localized species



Bacia & Schwille, Nature Protocols, 2007



FCCS - Fluorescence Cross-Correlation Spectroscopy

For 1:1 binding between **red** & **green** species and **no cross-talk**

$$A_G = 1/(N_{PP.\text{free}} + N_{LPNP})$$

$$A_R = 1/(N_{\text{Lip.free}} + N_{LPNP})$$

$$A_X = N_{LPNP}/\left((N_{PP.\text{free}} + N_{LPNP})(N_{\text{Lip.free}} + N_{LPNP})\right)$$

Fraction of coated PPs

$$\frac{A_X}{A_R} = \frac{N_{LPNP}}{N_{LPNP} + N_{PP.\text{free}}}$$

Fraction of used lips.

$$\frac{A_X}{A_G} = \frac{N_{LPNP}}{N_{LPNP} + N_{\text{Lip.free}}}$$

Relative values of the amplitudes $G(0)$ of **Green**, **Red**, and **Cross-correlation**

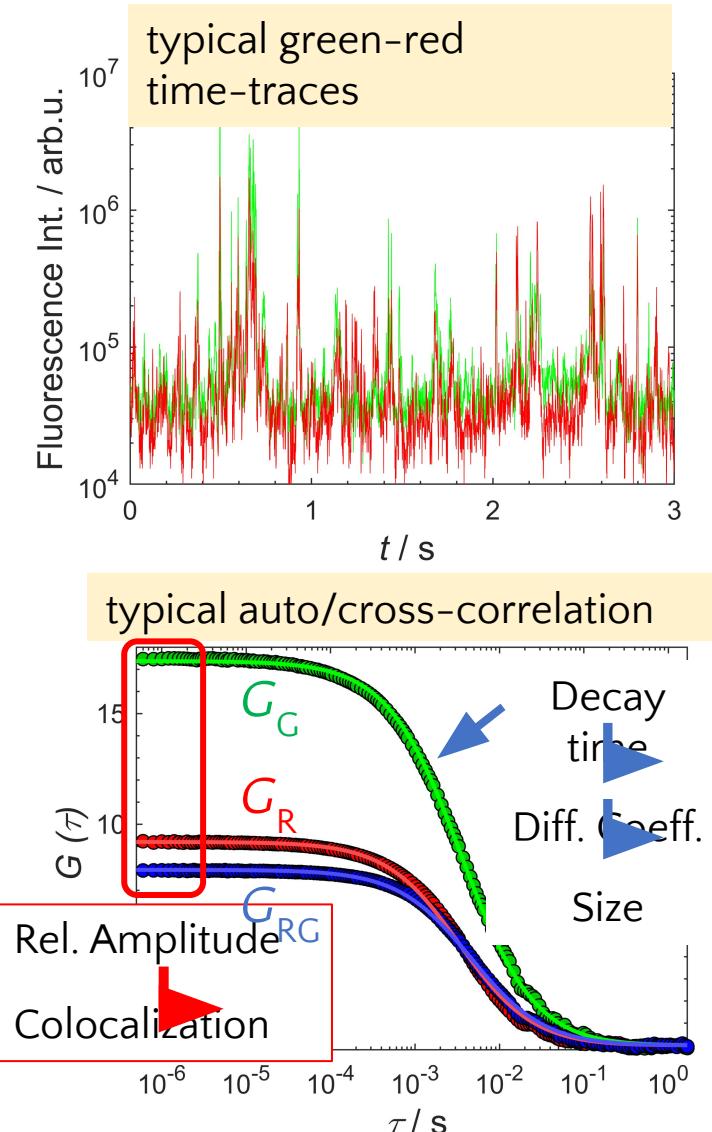
Extent of association between PPs and Lipos to form LPPs

PLL labeled with
Atto 488-NHS

Liposomes labeled
with Texas-RED

LPNP

Bacia & Schwille, Nature Protocols, 2007



FCCS quantifies colocalization between Lipos. & cores

Labels:

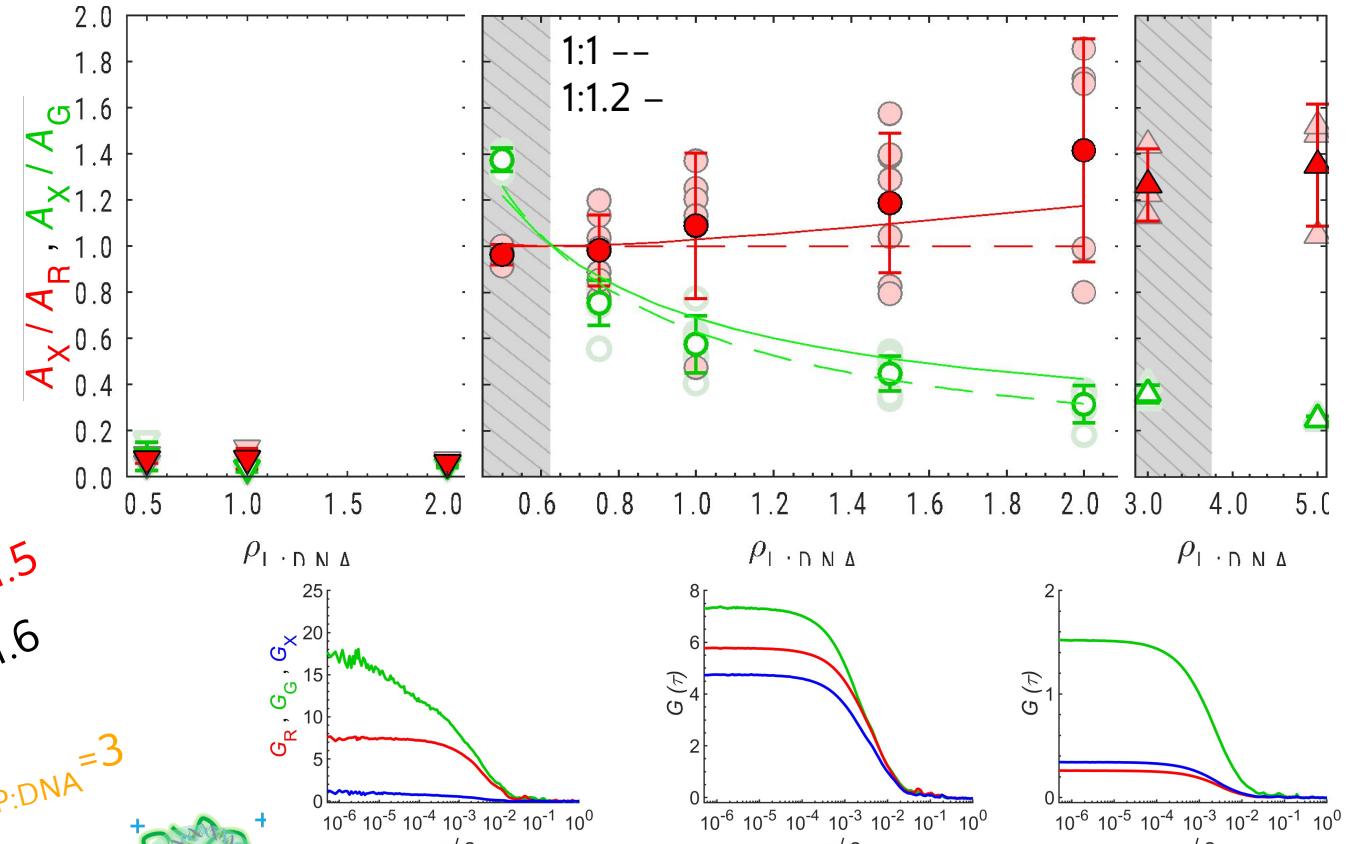
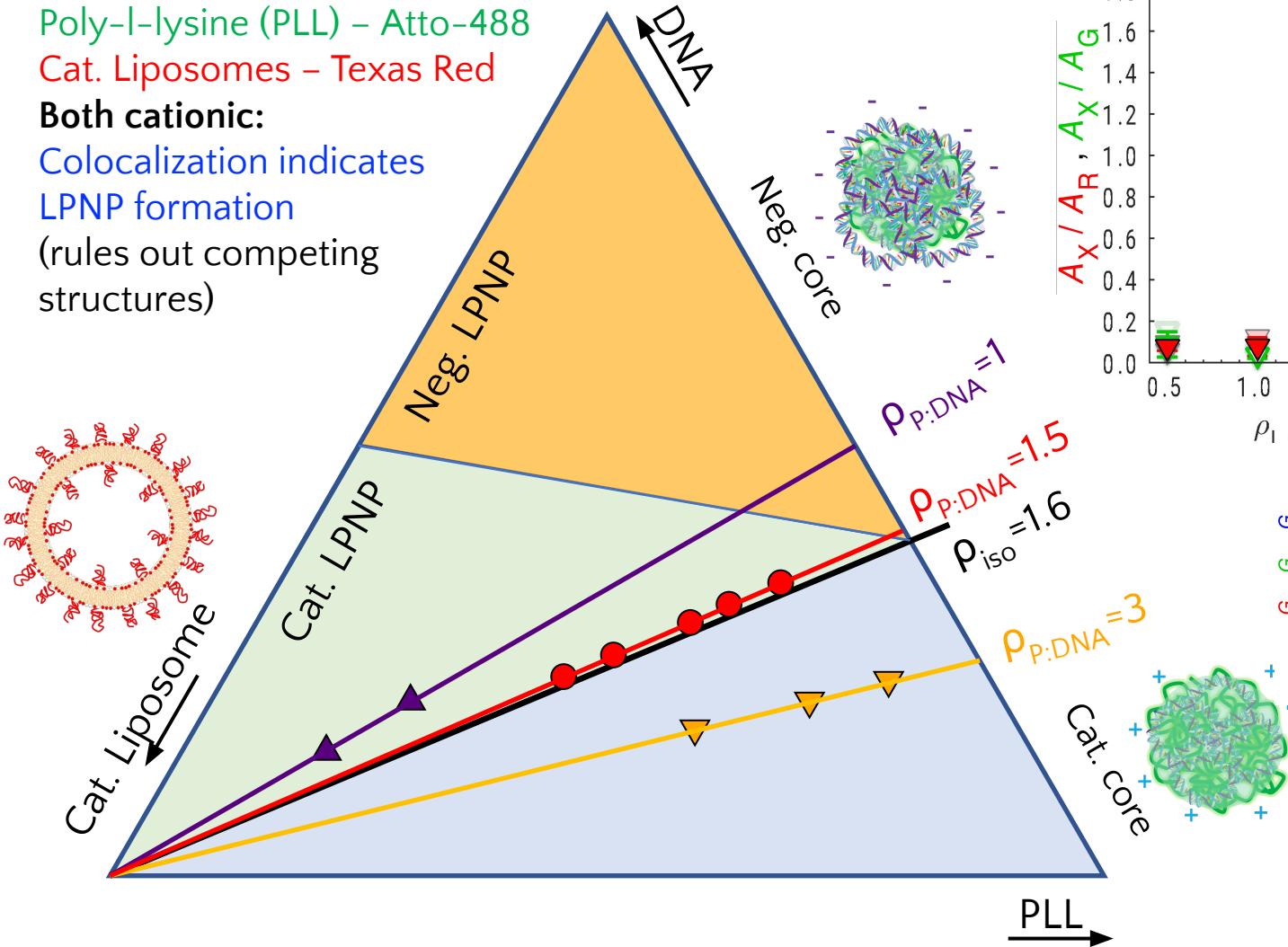
Poly-L-lysine (PLL) – Atto-488

Cat. Liposomes – Texas Red

Both cationic:

Colocalization in LPNP formation

(rules out competing structures)



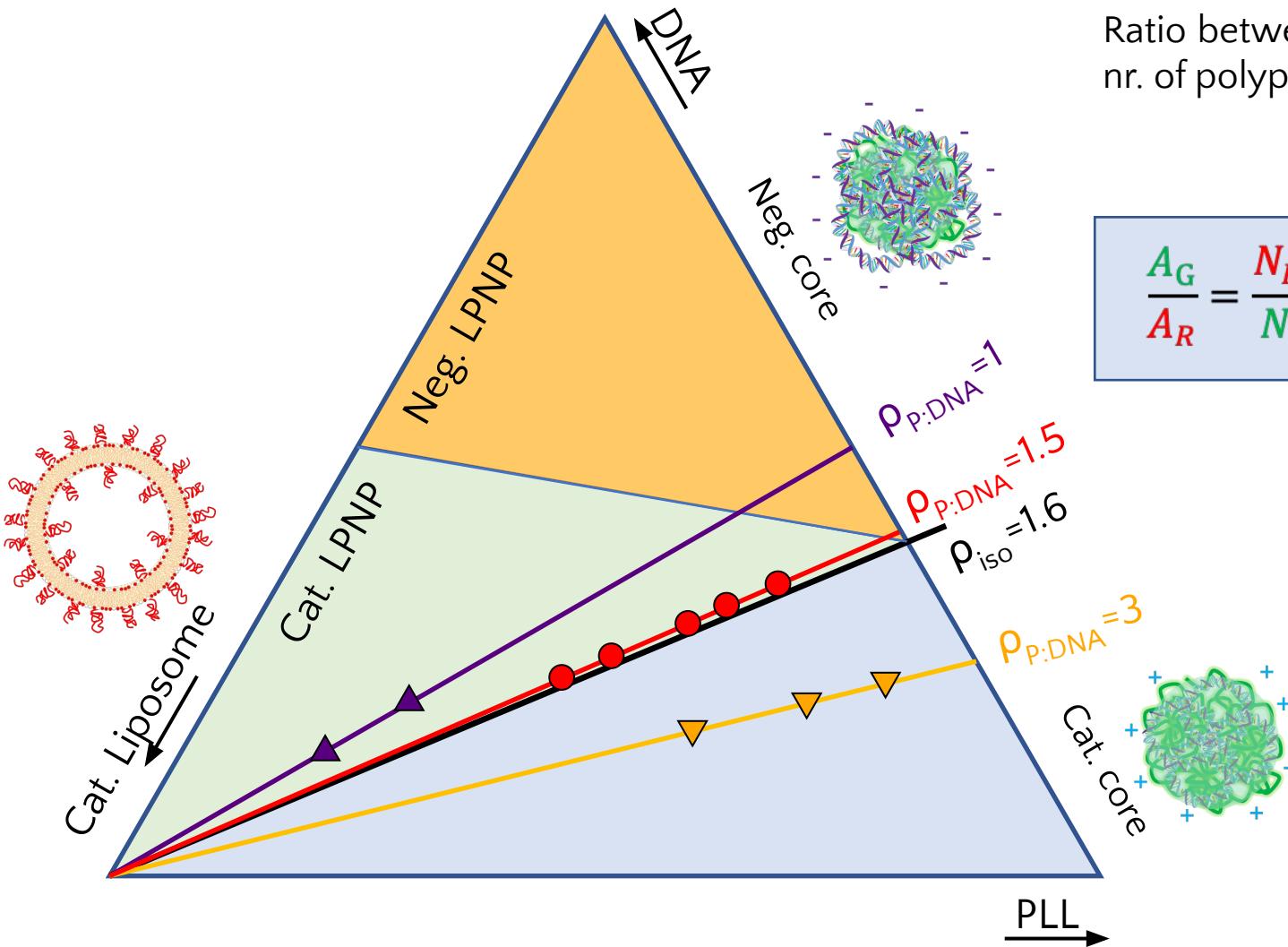
Fraction of coated PPs

$$\frac{A_X}{A_R} \approx \frac{N_{\text{LPNP}}}{N_{\text{LPNP}} + N_{PP,\text{free}}}$$

Fraction of used liposomes

$$\frac{A_X}{A_G} \approx \frac{N_{LPNP}}{N_{LPNP} + N_{Lip.\,free}}$$

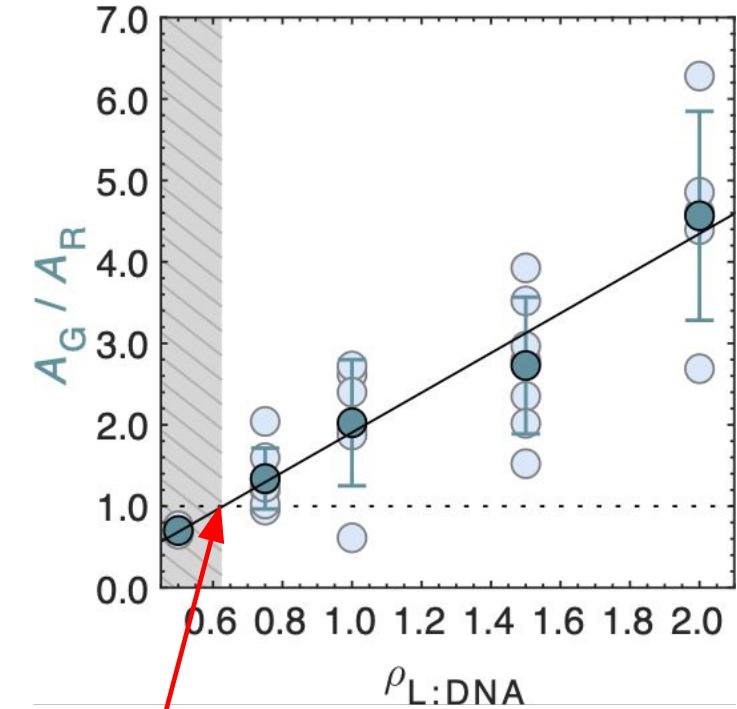
FCCS identifies critical ratio between nr of cores & lipos.



Since only slight deviation from 1:1 stoichiometry ($n < 1.5$):

Ratio between total nr. of liposomes (Liposomes + LPNPs) and total nr. of polyplexes (PPs + LPNPs) can be obtained from the A_G/A_R ratio

$$\frac{A_G}{A_R} = \frac{N_{Lip.Total}}{N_{PP.Total}}$$



Colloidal stability occurs when:
Liposomes to PPs nr ratio is at least 1 ($\rho_N \geq 1$)

FCCS identifies critical ratio between nr of cores & lipos.

Labels:

Poly-L-lysine (PLL) – Atto-488

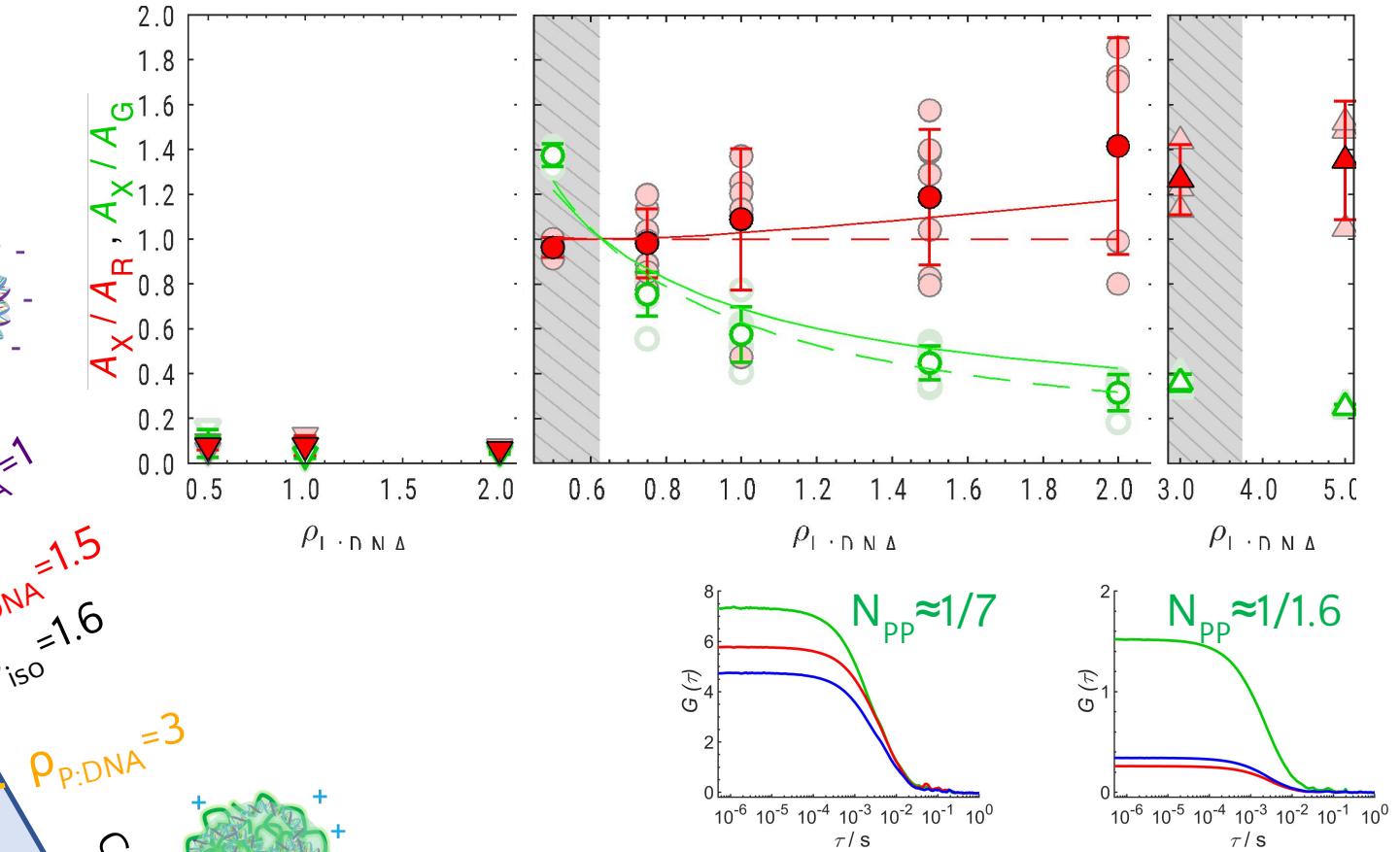
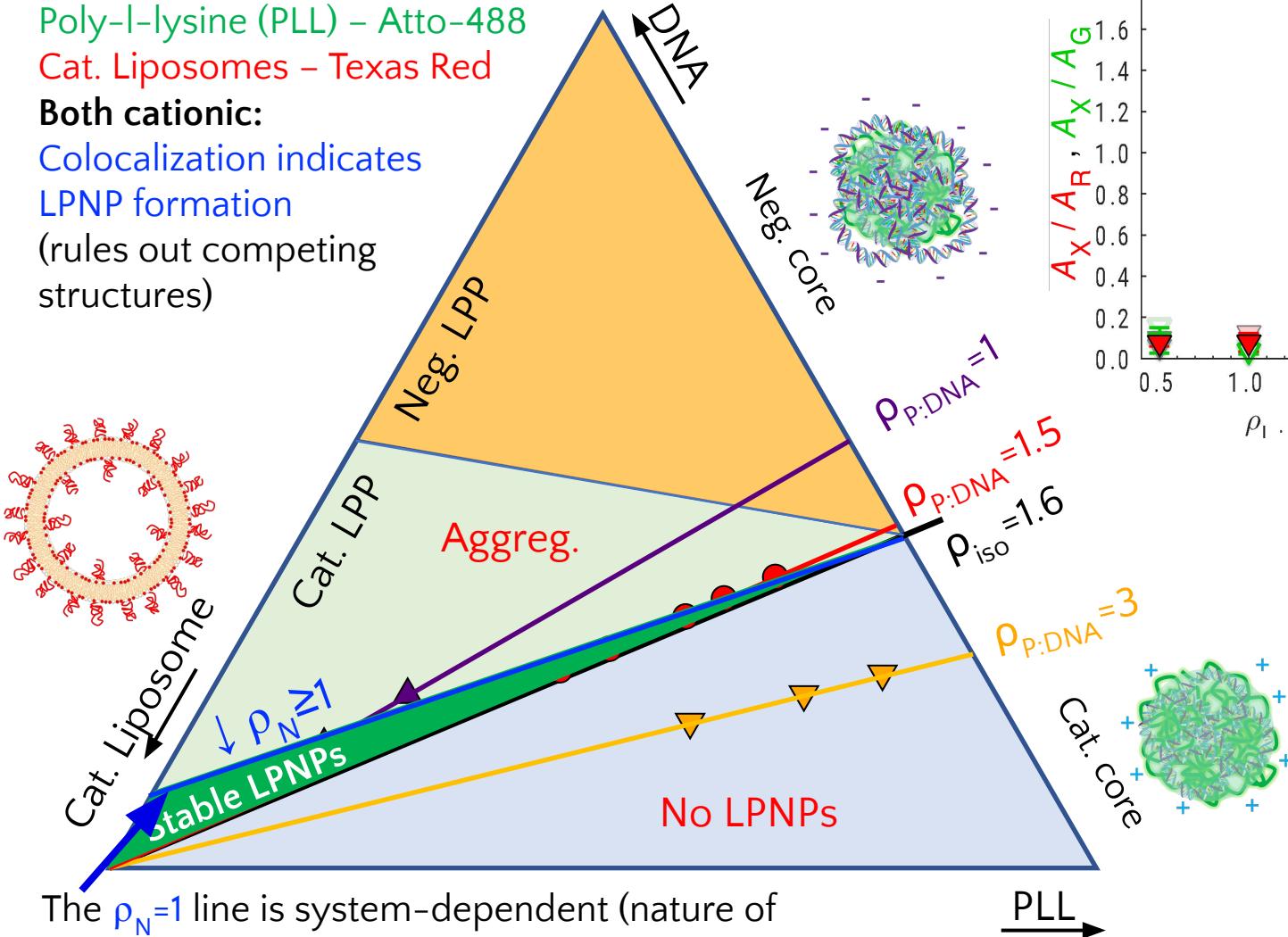
Cat. Liposomes – Texas Red

Both cationic:

Colocalization indicates

LPP formation

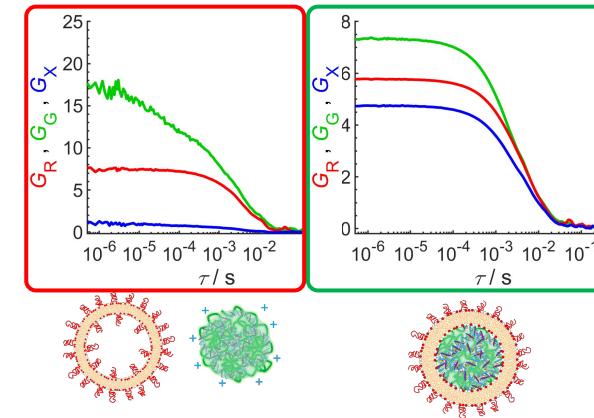
(rules out competing structures)



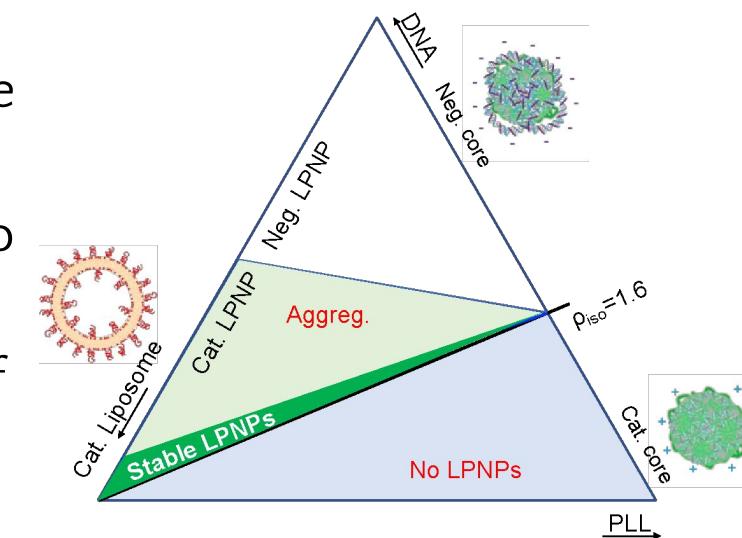
The nr. of PPs is ca. 3.5 times greater for $\rho_{P:DNA} = 1$, requiring also more liposomes to fulfill $\rho_N \geq 1$ criteria

Conclusions

- FCCS is a **powerful** tool for hybrid LPNPs characterization
 - **quantitative information** about the **core-shell association** (difficult to obtain by traditional techniques)
 - **insight** on forces/factors influencing association – LPNP synthesis optimization



- For Polylysine LPNPs
 - Association always between oppositely-charged particles (in simple buffer)
 - The liposome to polyplex number ratio **must** be at least 1 ($\rho_N \geq 1$) to ensure every core is coated and avoid aggregation
 - Polyplex (core) with **slight overcharge** – to avoid coexistence of species



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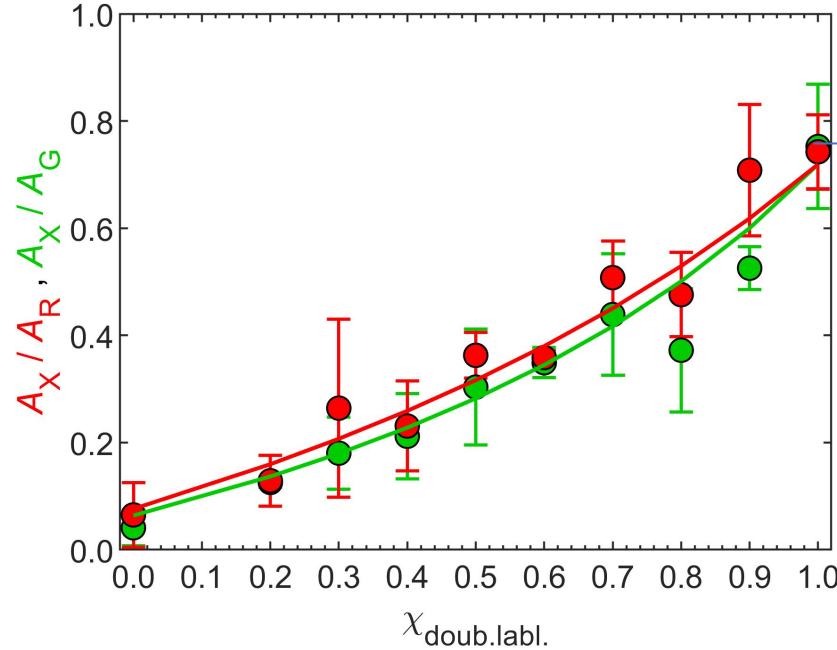
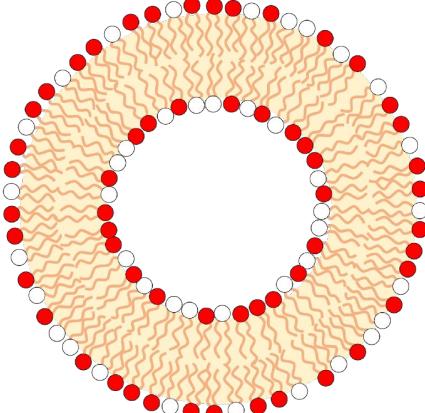
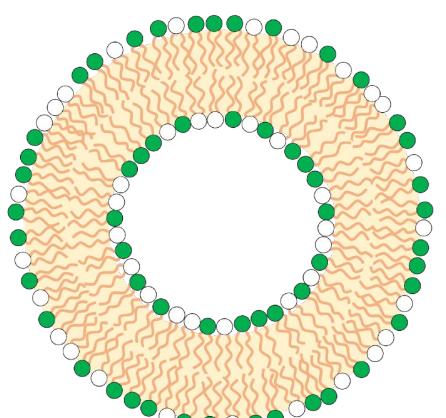
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Thank you for your Attention

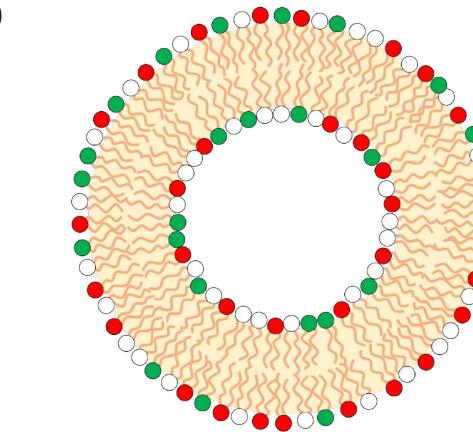
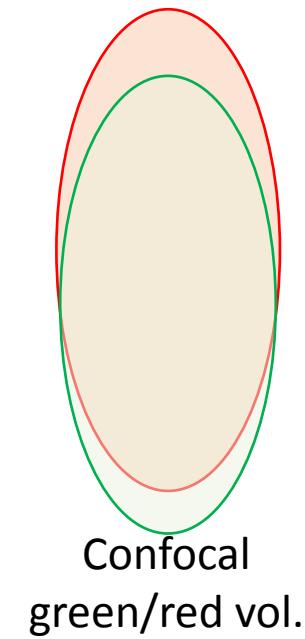
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FCCS: control measurements and volume overlap



Overlap volume
correction: **72%**



Similar to: Werner, Ebenhan, Haupt, Bacia, ChemPhysChem, 2018, 19, 3436