

# Dissolving Microarray Patches for Malaria Chemoprophylaxis:

*Mapping solid drug nanoparticle incorporation and long-acting release in vivo.*

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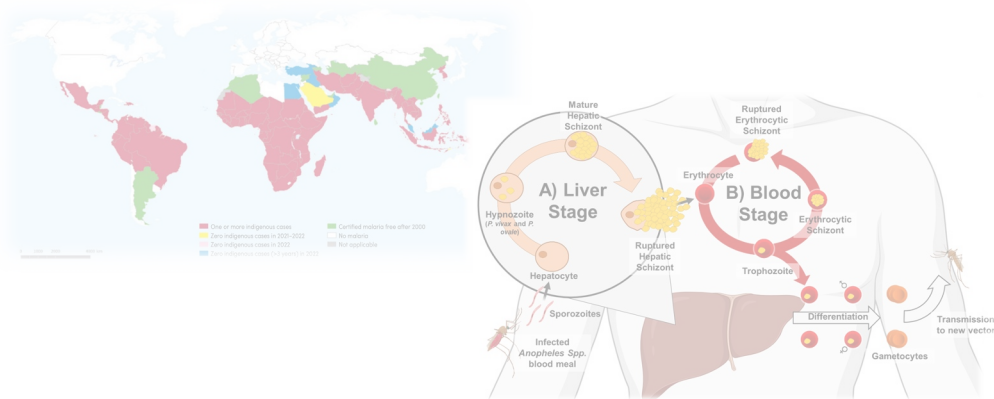


# The Malaria Burden

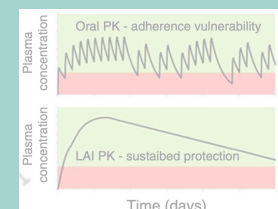
Parasitic infection caused by *Plasmodium* spp. - exclusively transmitted by female *Anopheles* spp. mosquitos.

Yearly Statistics (2022):<sup>1</sup>

- **249,000,000** new infections.
- **608,000** deaths.
- **76%** of deaths occur in children under five years old.
- **Every second**, a child in Africa dies from malaria.

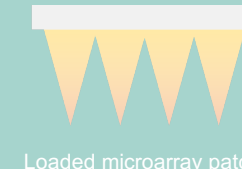
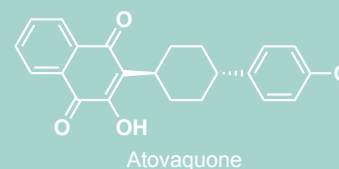


Previous work: Long-acting injectable formulation of atovaquone solid drug nanoparticles.<sup>2</sup>



*How might we improve adherence further?*

Long-acting antimalarial patch based on combined nanoformulation and microneedle technologies.



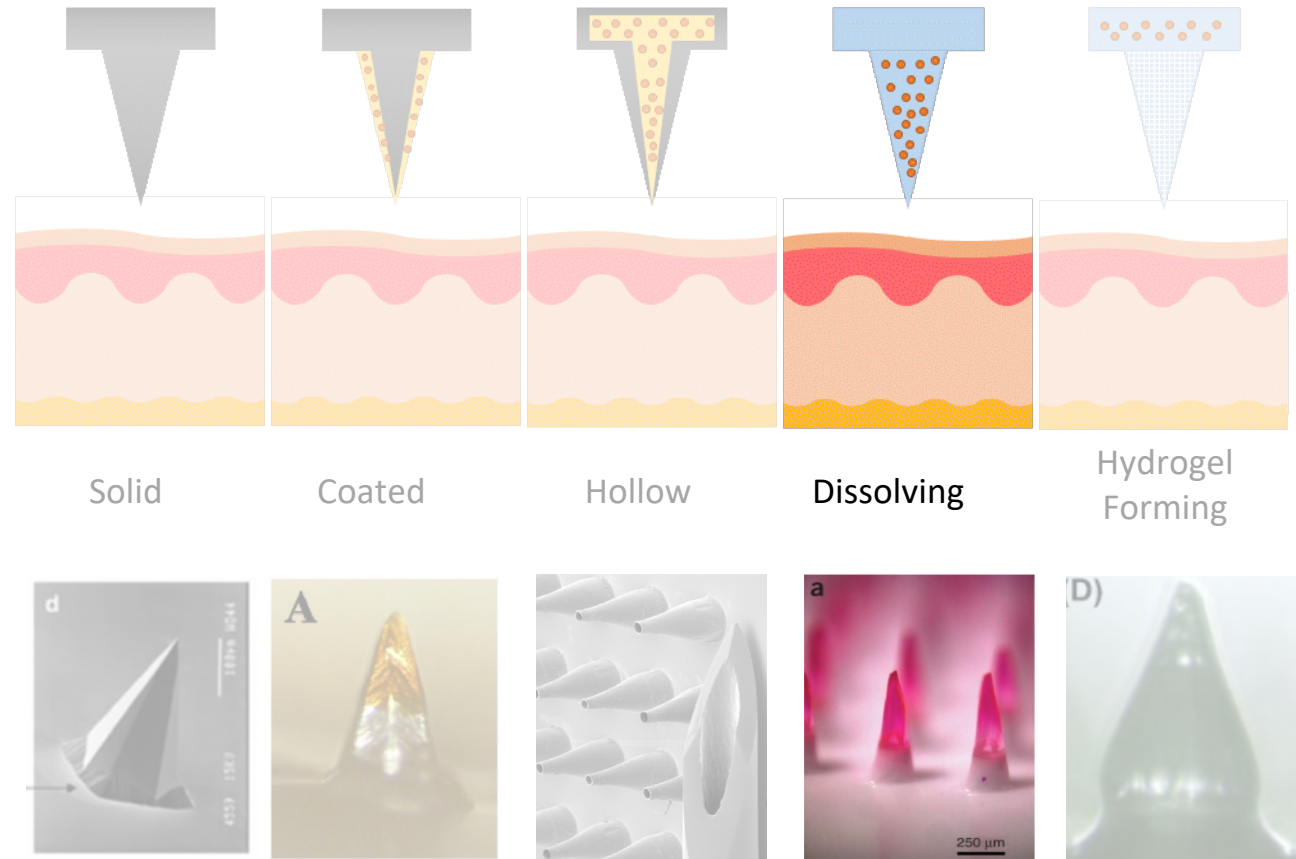
[1] World Health Organization, *World Malaria Report 2023*, 2023

[2] R.P. Bakshi *et al.*, *Nat. Commun.*, 2018, **9**, 315



# Microneedles

- Microneedles (MNs) as a Drug-Delivery System (DDS)
  - **Sub-millimetre** scale
  - Multiple MN architectures
  - **Enhance permeation** across a biological barrier (e.g. skin, cornea) while maintaining a **minimally-invasive** and **pain-free** profile.
- Dissolving Microneedles (DMNs)
  - Also known as dissolving microarray patches (DMAPs)
  - Pharmaceutical ingredient is incorporated within a water-soluble polymer matrix
  - Upon delivery, polymer dissolves to release the payload
  - Allows for circumvention of patient adherence issues such as **pill fatigue** and **needle phobia**
  - Suitable for deployment in **low-income areas**, dissolution removes the need for functional **sharps waste streams** associated with other MN technologies.



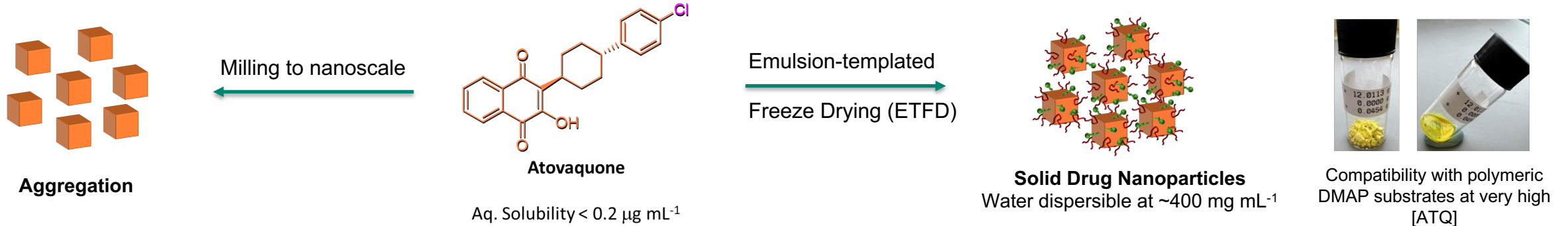
Images courtesy of:

E. Larraneta *et al.*, *Pharm Res*, 2016, **33**, 1055-1073  
 J Chen *et al.*, *Drug Dev. Ind. Pharm.*, 2013, **41**, 415-422  
 S. P. Davis *et al.*, *IEEE Trans. Biomed. Eng.*, 2005, **52**, 909-15  
 S.P. Sullivan *et al.*, *Nature Medicine*, 2010, **16**, 915-920  
 R. E. Lutton *et al.*, *Int. J. Pharm.*, 2015, **494**, 417-429

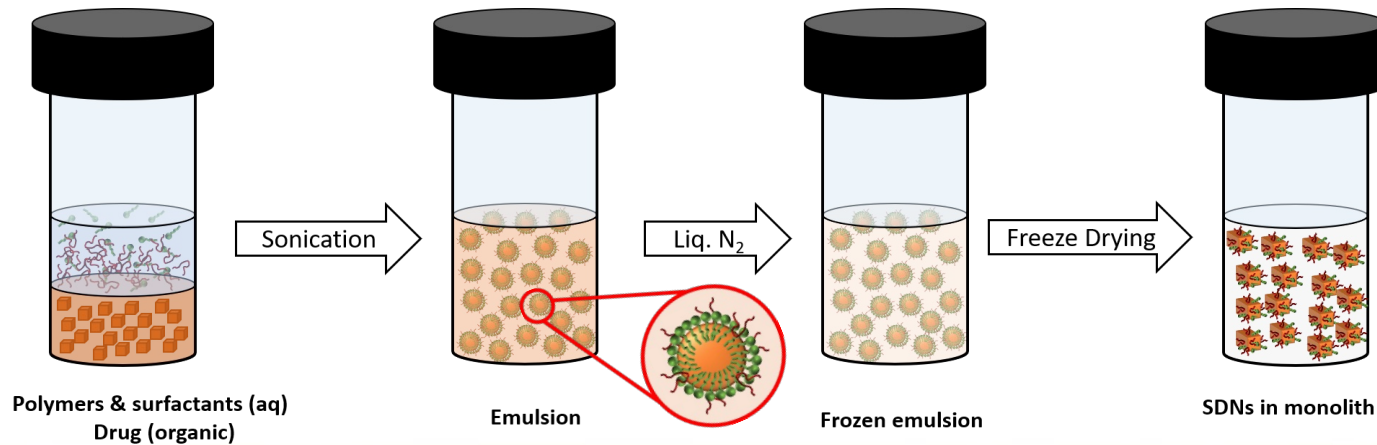


# Formulation: Solid Drug Nanoparticles

Atovaquone (ATQ) exhibits very poor aqueous solubility ( $< 1 \text{ mg mL}^{-1}$ ). Its incorporation into a water-soluble DMAP therefore requires nanoformulation.



## Nanoformulation approach: Emulsion-templated Freeze-drying (ETFD)



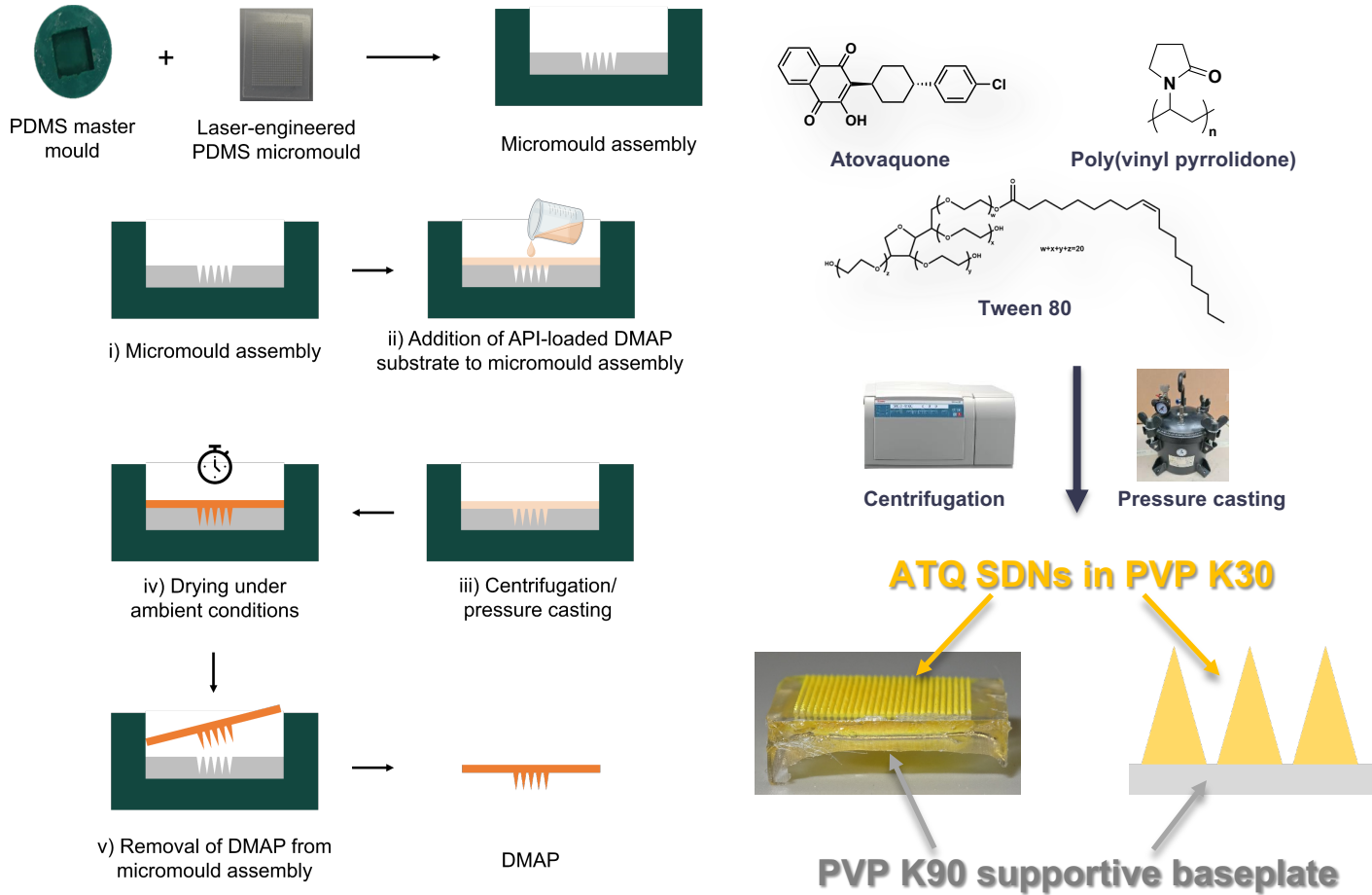
- 1) Generation of oil-in-water mixture of the drug (oil) and excipient stabilisers (water).
- 2) Energetic emulsification to form nanoscale oil droplets, stabilized by amphiphilic polymer and surfactant stabilisers (excipients).
- 3) Cryogenic freezing of the emulsion, trapping the stabilized oil droplets within a matrix of ice containing excess polymeric excipients.
- 4) Lyophilisation of the frozen emulsion. Solvents (water and organic) are removed leaving a porous polymer matrix with drug nanoparticles dispersed throughout.



# Formulation: Dissolving Microarray Patches

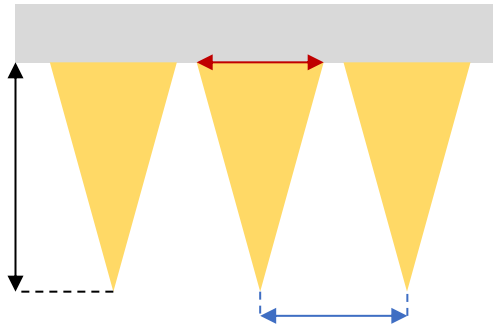
ATQ SDN-loaded DMAPs formulated using a solvent casting micromoulding approach using a highly concentrated suspension of ATQ SDNs dispersed in a polymeric matrix. Poly(vinyl pyrrolidone) was used as the DMAP substrate.

## Solvent casting micromoulding



# DMAPs – Important Characteristics

## MN Dimensions



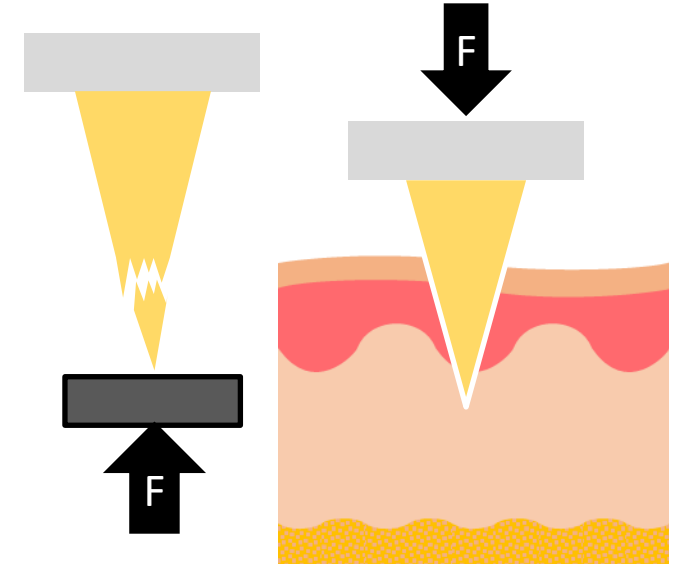
- MN height
  - Delivery of API to dermal microcirculatory system
- **Base width**
  - Insertion force *via* apex angle
- Interspacing
  - Insertion force *via* bed of nails effect

## API Distribution



- Homogeneity of drug loading
- API concentration at microneedle tips preferable
- Possible effects of API distribution on mechanical properties?

## Mechanical Properties



- MN integrity under compression
- Resistance to compression force
- Insertion force



# DMAP Dimension Analysis

DMAP Dimensions should be controlled by the CAD design of the laser-etched micromould:

Microneedle (MN) height = 600  $\mu\text{m}$

Microneedle base width = 300  $\mu\text{m}$ .

Microarray interspacing = 450  $\mu\text{m}$ .

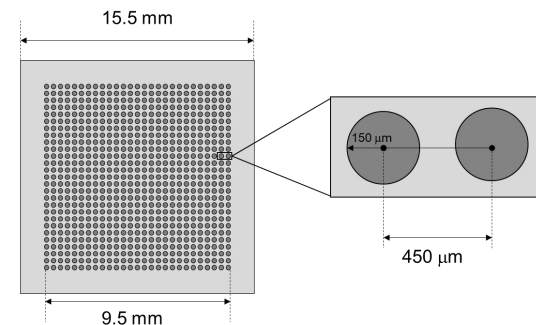


Fig. 1: CAD schematic of DMAP micromould.

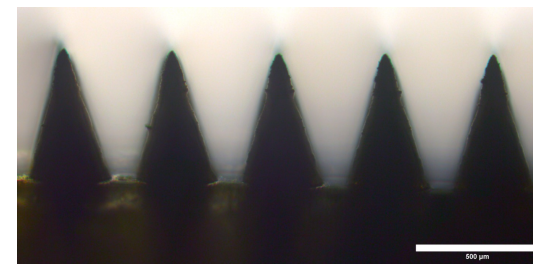
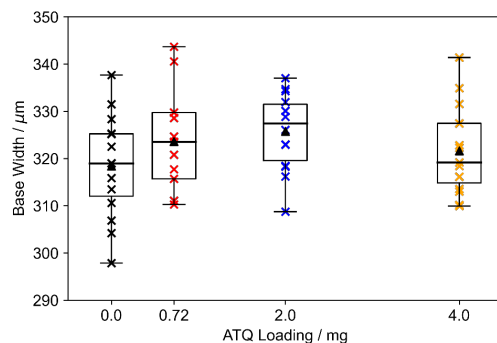
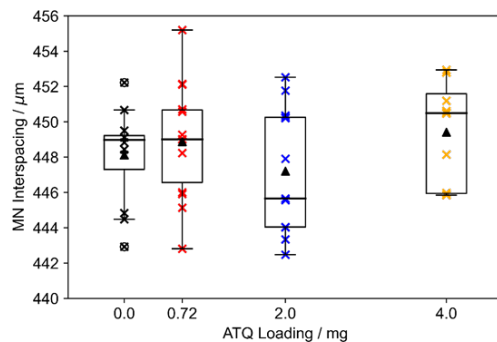


Fig. 2: Optical Microscope image of ATQ SDN-loaded DMAP.

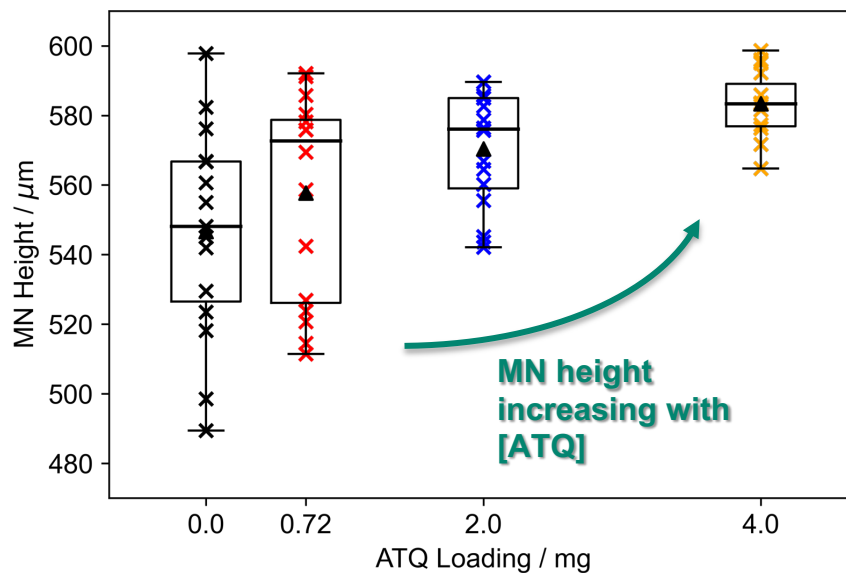
## Base Width via Optical microscopy



## Interspacing via Optical Microscopy



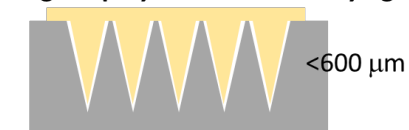
## MN Height via Optical Microscopy



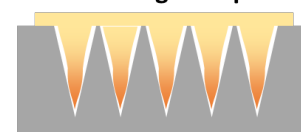
DMAP in micromould before drying



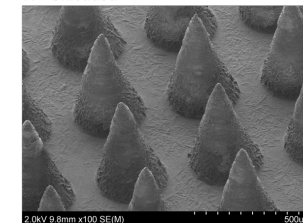
Shrinkage of polymer matrix on drying



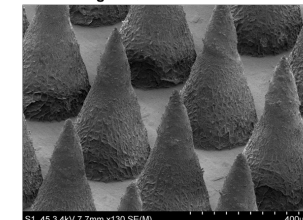
Asymmetric shrinkage of tip-loaded DMAPs



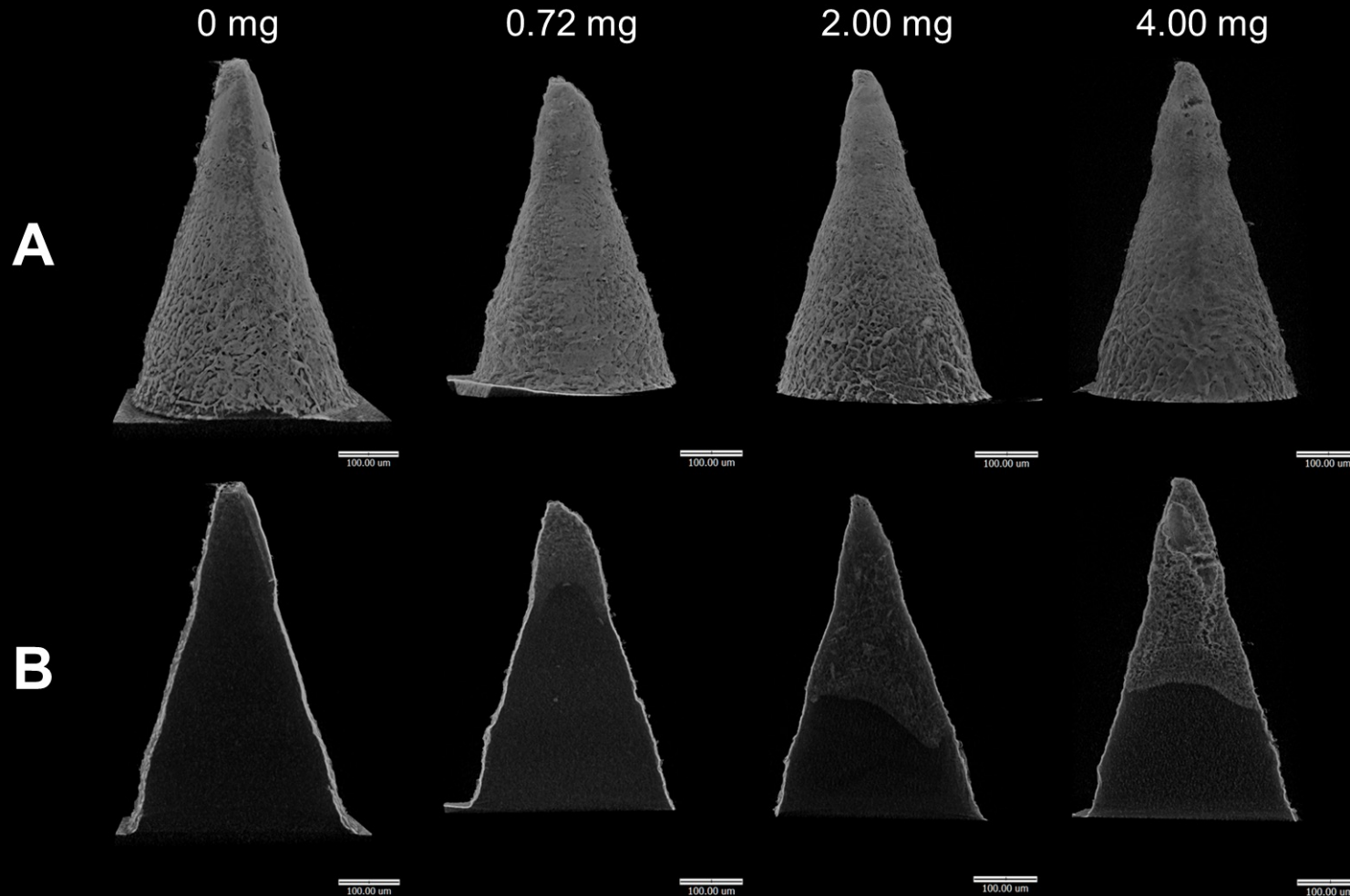
A – Unloaded



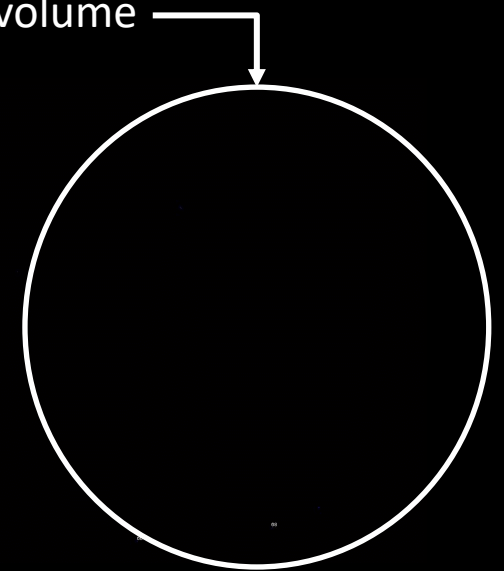
B – 0.72 mg ATQ



# API Distribution – X-ray Computerised Microtomography ( $\mu$ CT)



- Contrast = density
- Increased density at MN tips implies ATQ tip distribution
- Voids in 4.00 mg DMAP – inefficient packing
  - Voids are *ca.* 8% of MN volume



Object map of 4.00 mg DMAP





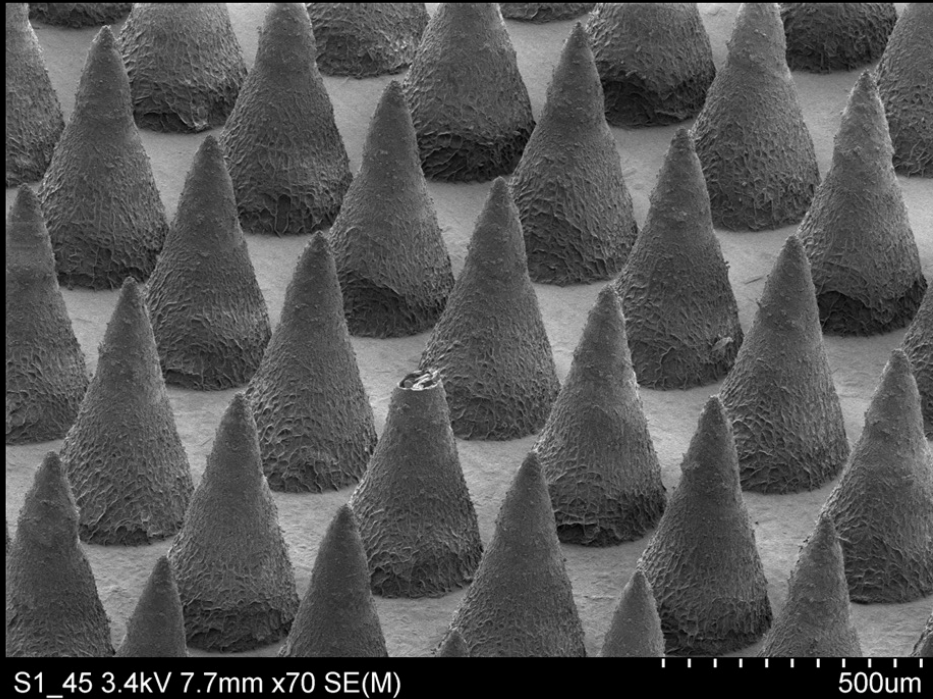
# API Distribution — Scanning Electron Microscopy/Energy Dispersive X-ray spectroscopy

Scanning Electron Microscopy (SEM) revealed broken MN tip...

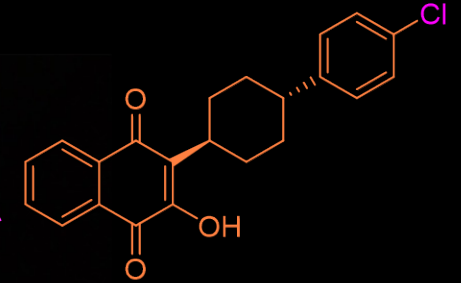
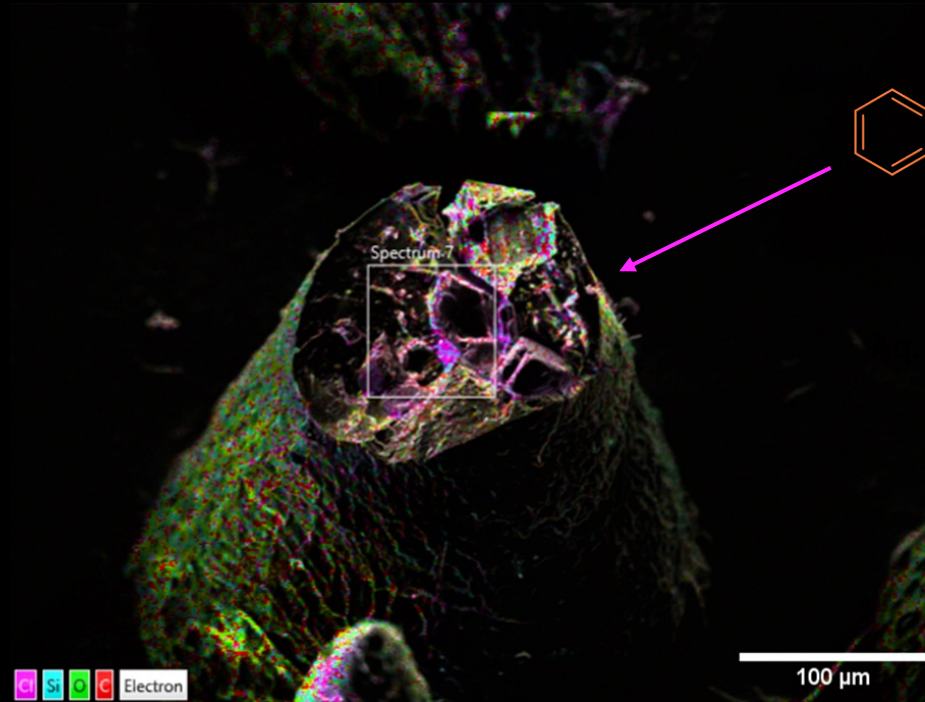
Energy-dispersive X-ray Spectroscopy (EDXS): Element mapping to SEM image

ATQ: C<sub>22</sub>H<sub>19</sub>ClO<sub>3</sub>

A) SEM image of 0.72 mg ATQ DMAP (with broken tip)



B) EDXS of broken tip

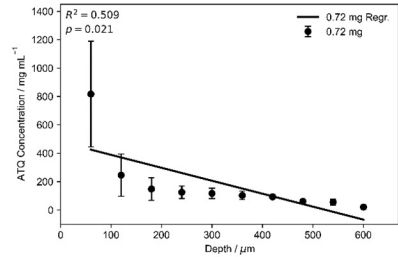
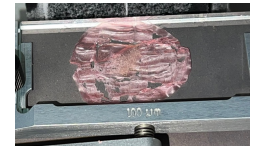


# API Distribution – Radiometric Sectioning

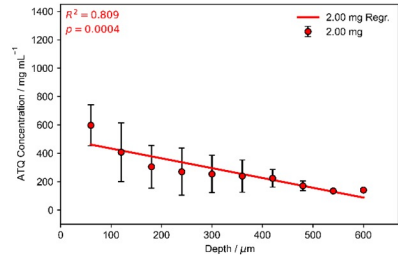


$^3\text{H}$ -ATQ SDN-loaded DMAPs embedded in dental wax and sectioned *via* cryostat.

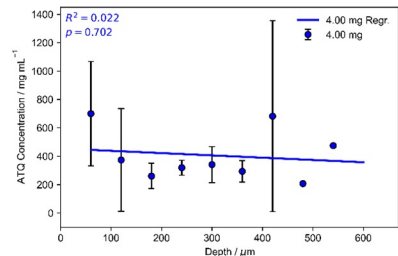
- Spatial  $^3\text{H}$ -ATQ distribution profiles obtained *via* radiometric mass balance.



0.72 mg ATQ



2.00 mg ATQ



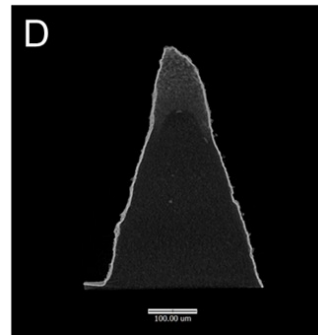
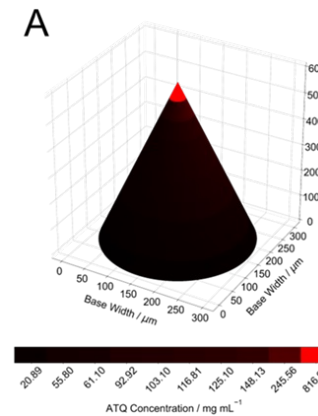
4.00 mg ATQ

- ATQ concentrated primarily in tip
- Gradual distribution thereafter

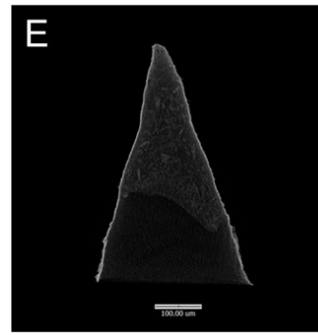
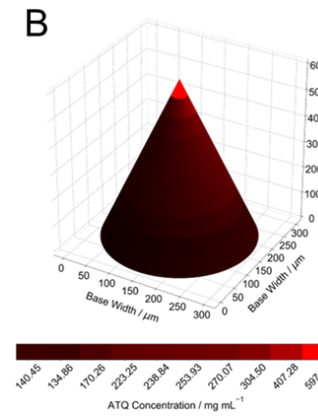
- Less dramatic tip concentration
- More even distribution of ATQ across MNs

- Gradual distribution of ATQ
- Highest average ATQ concentration in tip
- Highly variable (voids?)

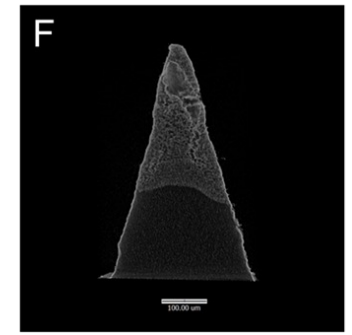
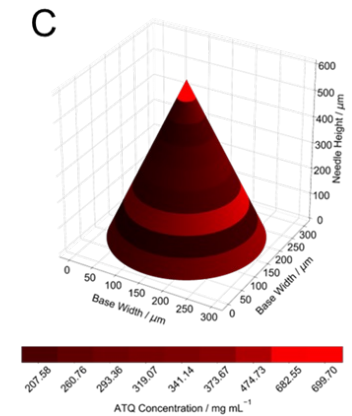
0.72 mg ATQ



2.00 mg ATQ



4.00 mg ATQ



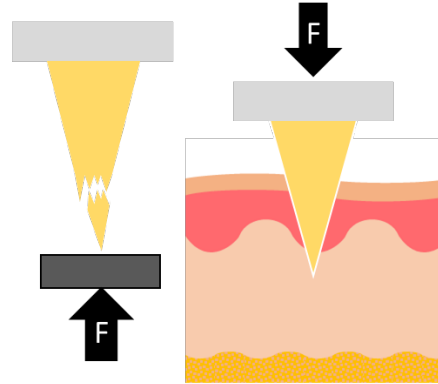


# DMAP Mechanical Properties

$$\text{Young's Modulus } (E) = \frac{\sigma \text{ (force per unit area)}}{\varepsilon \text{ (proportional deformation of sample)}} ; \text{ Needle Modulus} = \frac{\text{Force applied per needle } (N)}{\varepsilon}$$

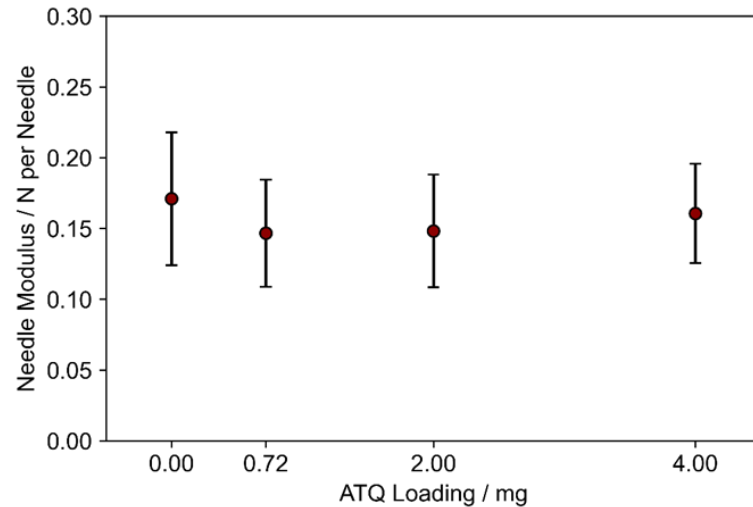
Resistance to axial compression did not vary with ATQ loading.

## Mechanical Properties

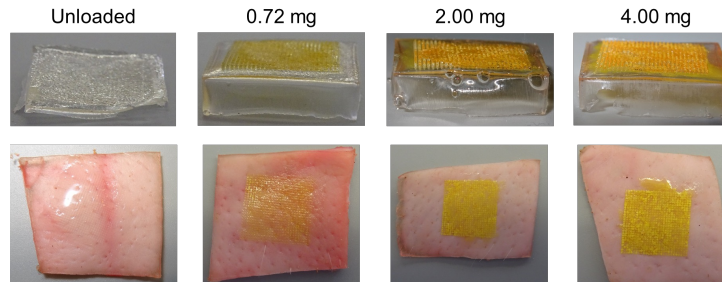


Complete DMAP insertion into ex vivo porcine skin attainable by patient-generable forces

Needle moduli of ATQ SDN-loaded DMAPs

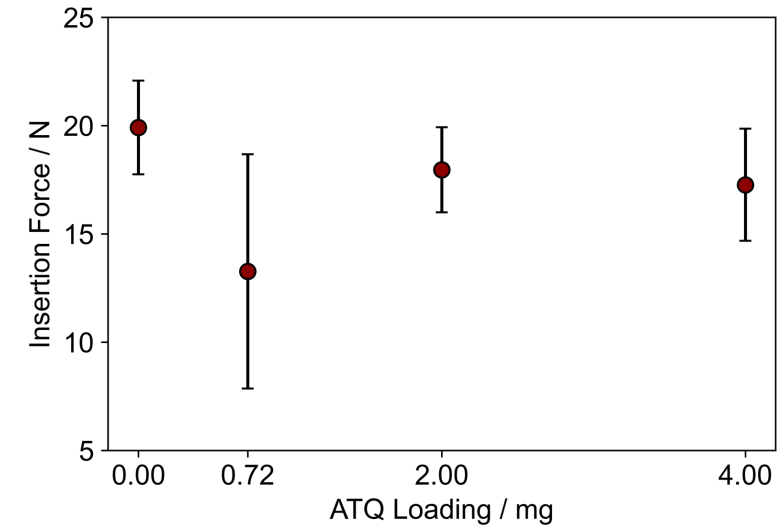


- MN integrity under compression
- Resistance to compression force
- Insertion force



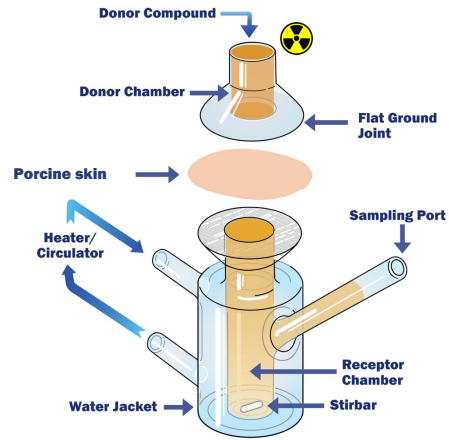
Dissolution and release of ATQ SDNs on ex vivo insertion

Ex vivo insertion force of ATQ SDN-loaded DMAPs

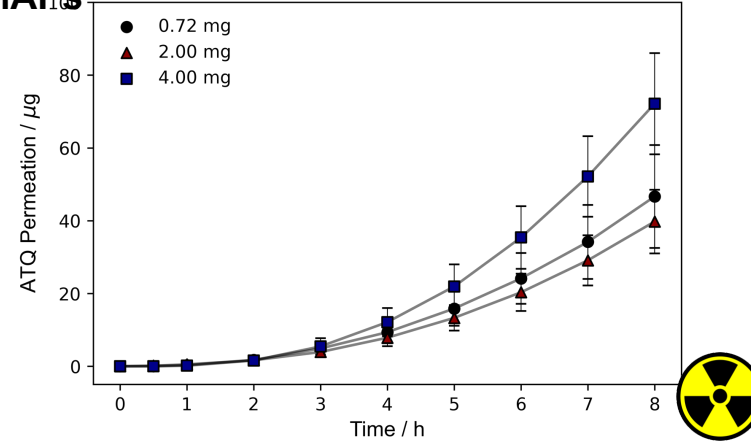


# Ex vivo/in vivo Release

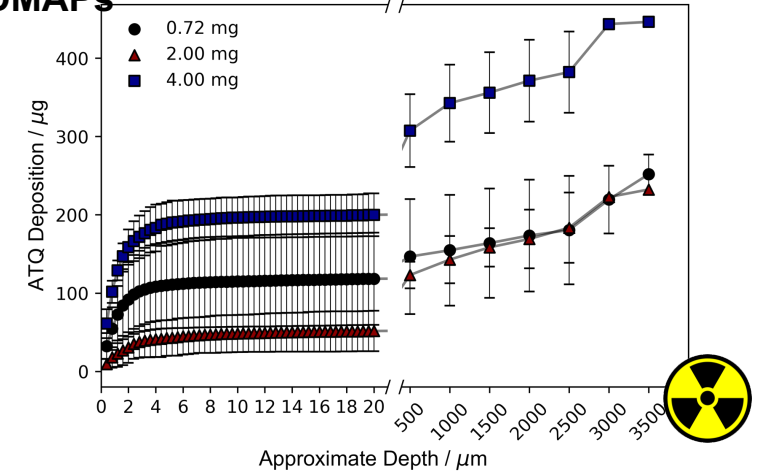
## Ex vivo Radiometric Permeation/Deposition



## Radiometric ex vivo permeation of ATQ from DMAPs



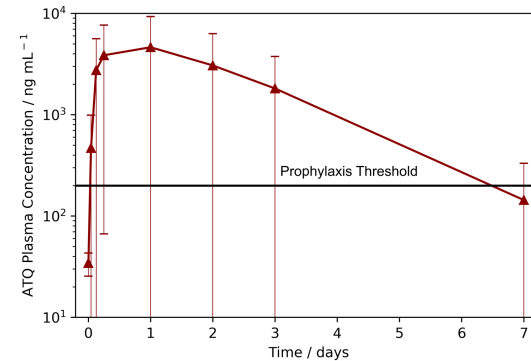
## Radiometric ex vivo deposition of ATQ from DMAPs



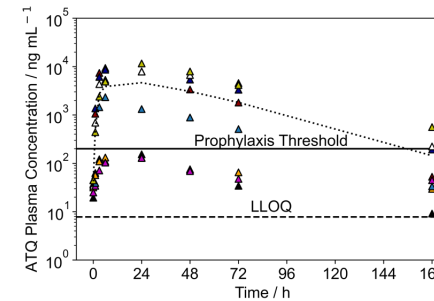
## In vivo Pharmacokinetics



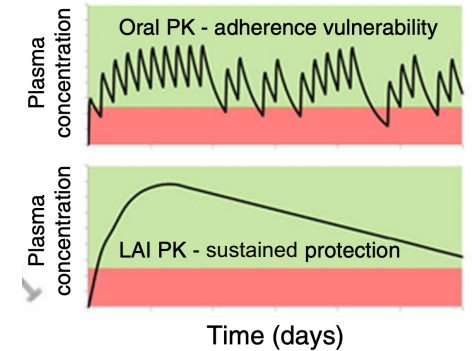
24-hour application  
of microarray  
patch...



...up to 7 days of predicted prophylaxis following a single patch application.



\*Response group deconvolution



Long-acting  
pharmacokinetics



# Conclusions

- Successful formulation of highly concentrated ATQ SDN-loaded DMAPs
- ATQ is spatially concentrated towards MN tips within SDN-loaded DMAPs
  - DMAP morphology is affected but mechanical properties are not.
- Ex vivo permeation/deposition studies may indicate depot formation and sustained release.
- ATQ SDN-loaded DMAPs show long-acting release of therapeutically relevant concentrations of ATQ *in vivo* following a single patch application.



# Acknowledgements

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